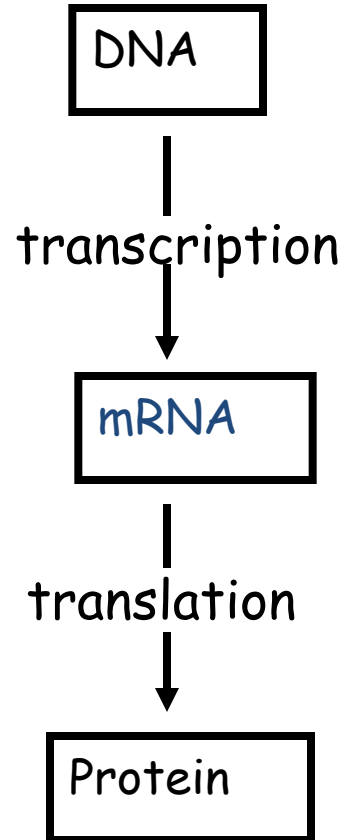
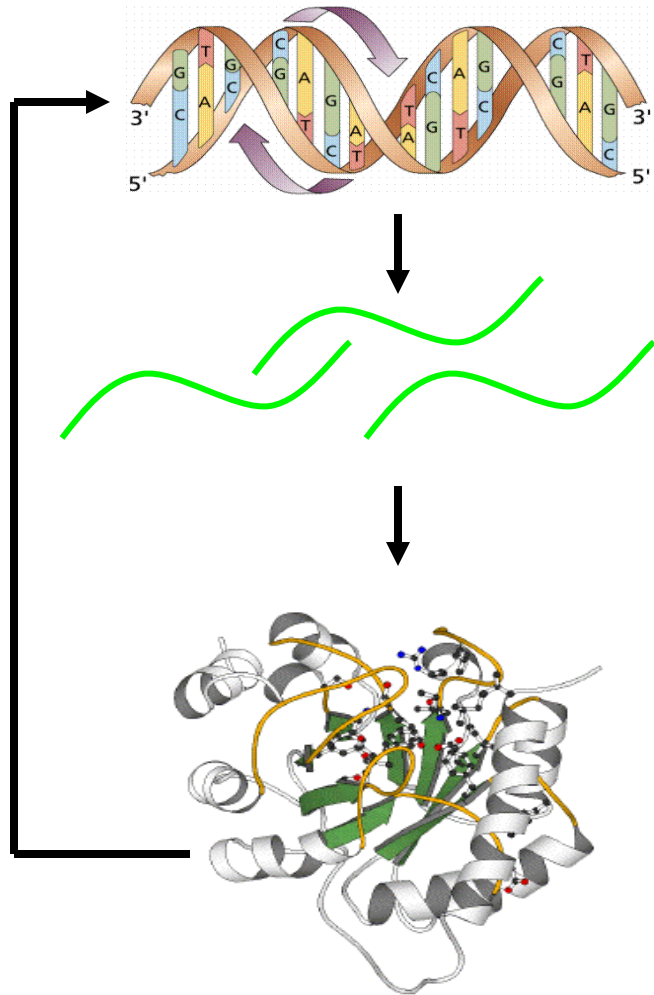


10-601
Machine Learning

HMM applications in computational
biology

Central dogma

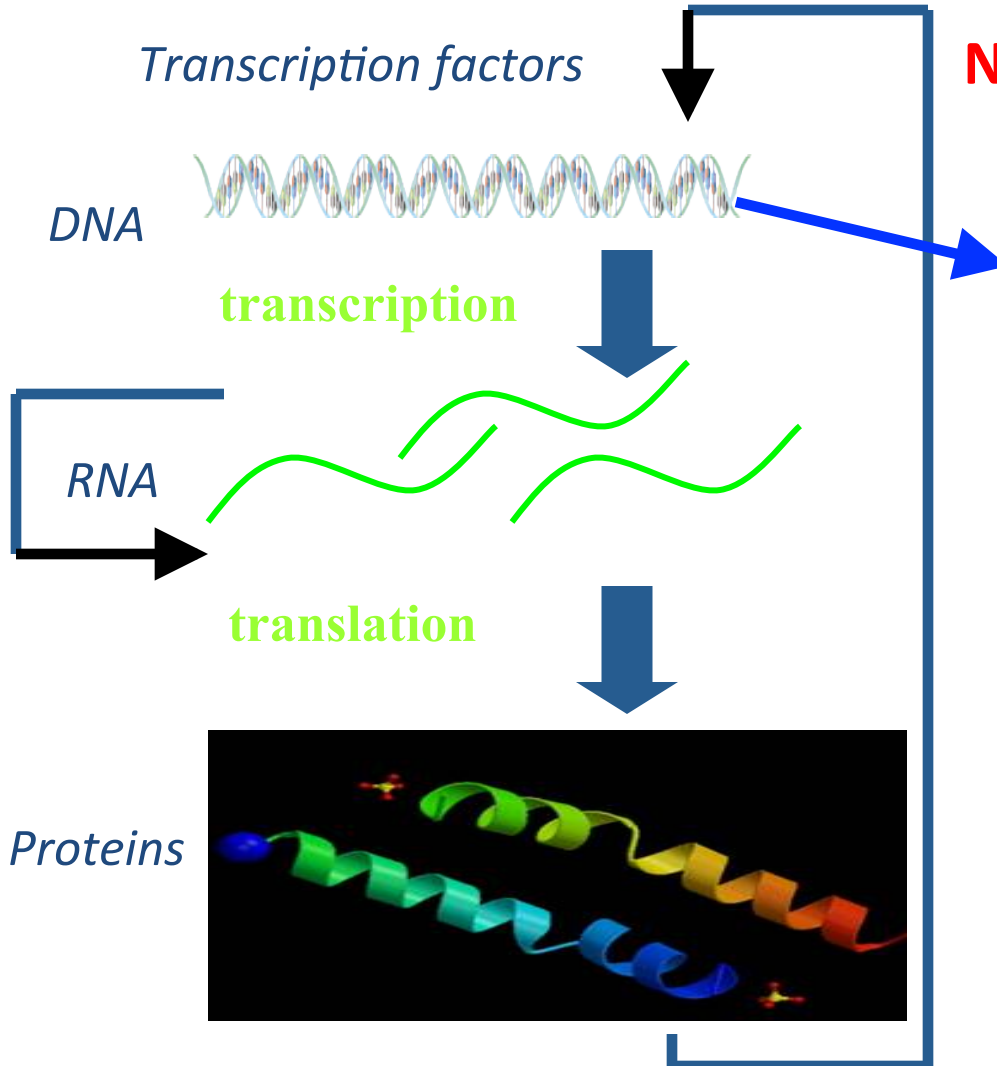


CCTGAGCCAAC TATTGATGAA

CCUGAGCCAACUAUUGAUGAA

PEPTIDE

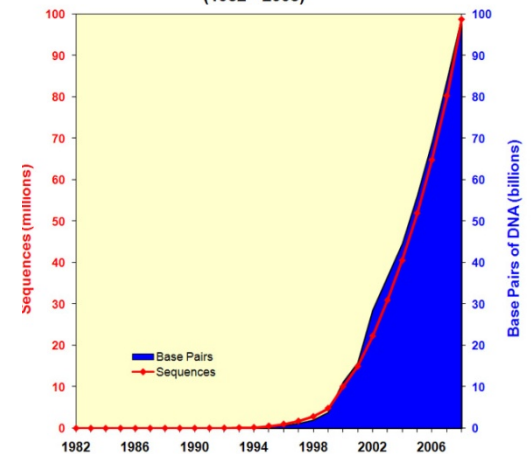
Biological data is rapidly accumulating



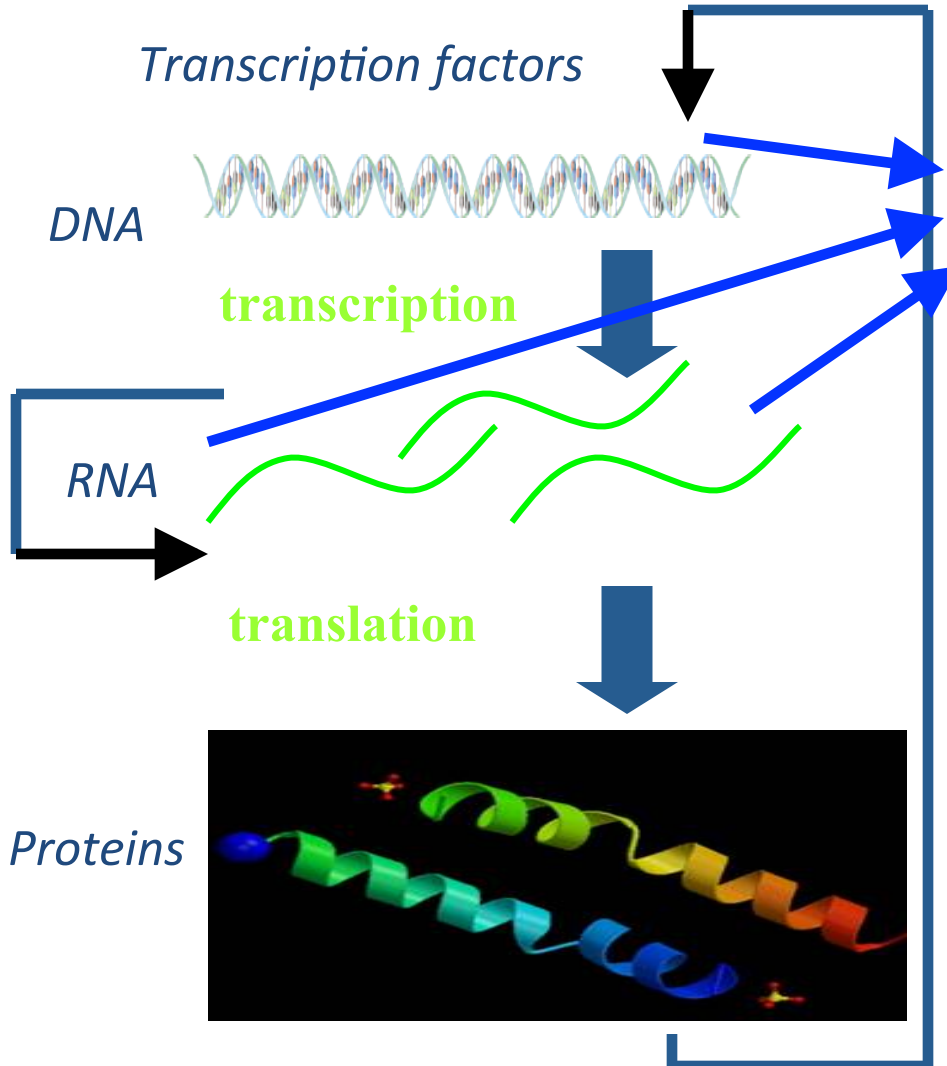
Next generation sequencing



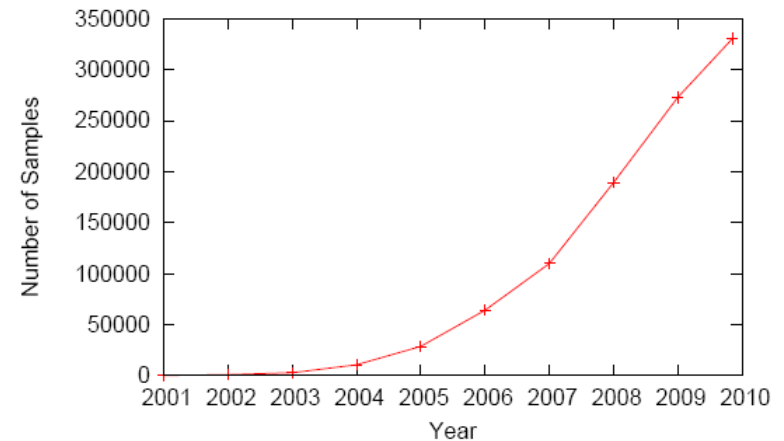
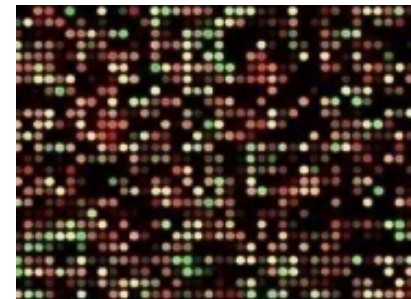
Growth of GenBank
(1982 - 2008)



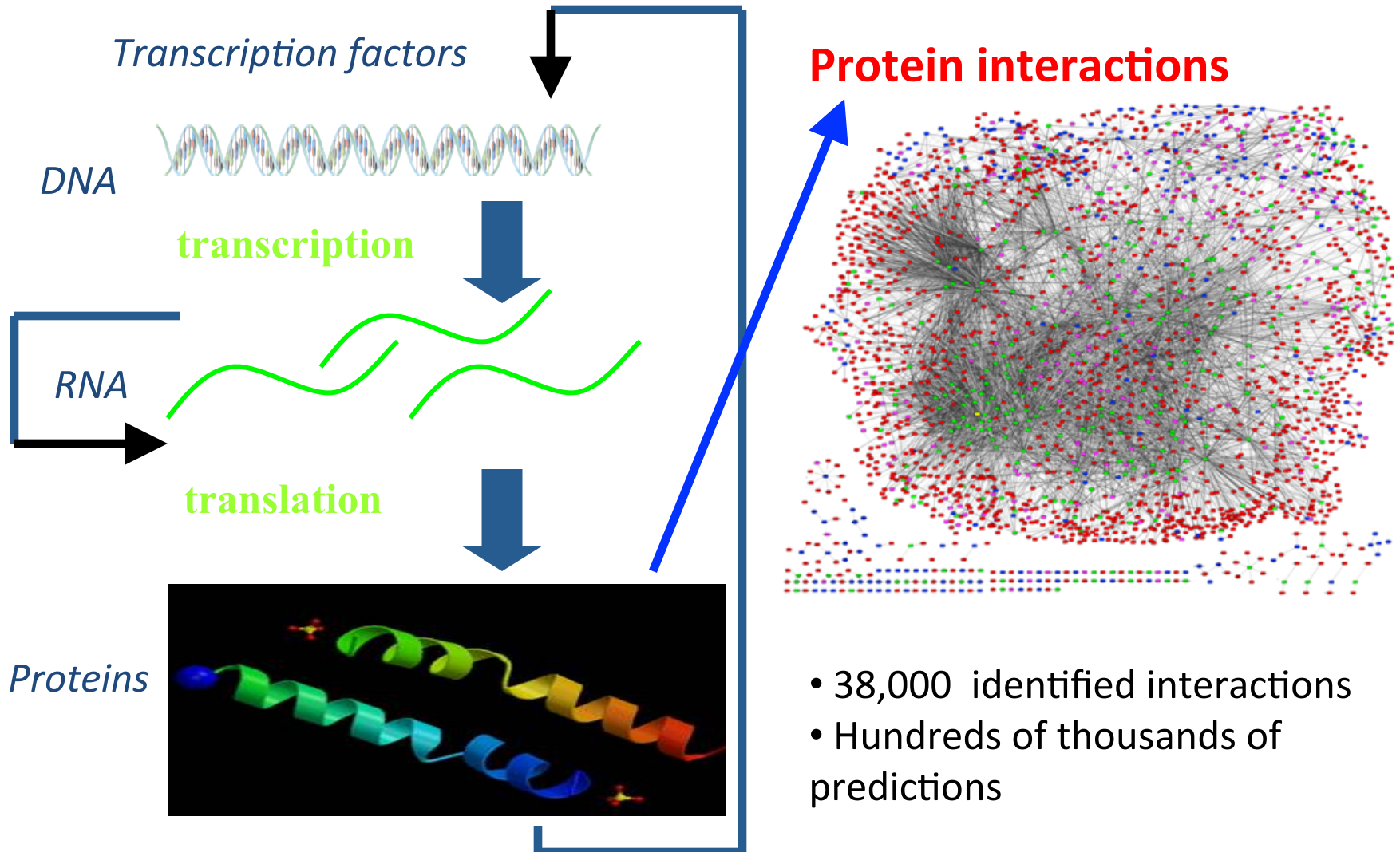
Biological data is rapidly accumulating



Array / sequencing technology



Biological data is rapidly accumulating



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Company Unveils DNA Sequencing Device Meant to Be Portable, Disposable and Cheap

By ANDREW POLLACK

Published: February 17, 2012

DNA sequencing is becoming both faster and cheaper. Now, it is also becoming tinier.

A British company said on Friday that by the end of the year it would begin selling a disposable gene sequencing device that is the size of a USB memory stick and plugs into a laptop computer to deliver its

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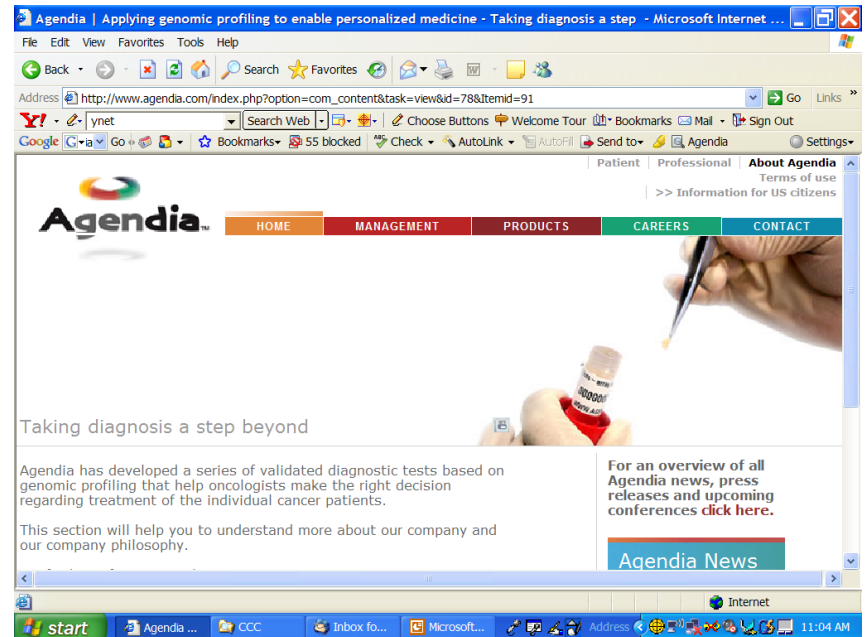
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FDA Approves Gene-Based Breast Cancer Test*

“ MammaPrint is a DNA microarray-based test that measures the activity of 70 genes in a sample of a woman's breast-cancer tumor and then uses a specific *formula* to determine whether the patient is deemed low risk or high risk for the spread of the cancer to another site.”



*Washington Post, 2/06/2007



Understand change with new tools for epigenetics research from BioLabs Inc.



New Software Section PLoS Computational Biology accepting presubmission inquiries

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Metabolic Factors Limiting Performance in Marathon Runners

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Benjamin I. Rapoport^{1,2*}

1 M.D.– Ph.D. Program, Harvard Medical School, Boston, Massachusetts, United States of America, **2** Department of Electrical Engineering and Computer Science and Division of Health Sciences and Technology, Massachusetts Institute of Technology, Cambridge, Massachusetts, United States of America

Abstract [Top](#)

Each year in the past three decades has seen hundreds of thousands of runners register to run a major marathon. Of those who attempt to race over the marathon distance of 26 miles and 385 yards (42.195 kilometers), more than two-fifths experience

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Letters to Nature

Nature **427**, 247-252 (15 January 2004) | doi:10.1038/nature02236; Received 24 July 2003; Accepted 14 November 2003

Functional genomic hypothesis generation and experimentation by a robot scientist

Ross D. King¹, Kenneth E. Whelan¹, Ffion M. Jones¹, Philip G. K. Reiser¹, Christopher H. Bryant², Stephen H. Muggleton³, Douglas B. Kell⁴ & Stephen G. Oliver⁵

1. Department of Computer Science, University of Wales, Aberystwyth SY23 3DB, UK
2. School of Computing, The Robert Gordon University, Aberdeen AB10 1FR, UK
3. Department of Computing, Imperial College, London SW7 2AZ, UK
4. Department of Chemistry, UMIST, P.O. Box 88, Manchester M60 1QD, UK

Sequencing DNA



First human genome draft in 2001



Due to *accumulated errors*, we could only reliably read at most **100-200 nucleotides**.

DARPA Shredder Challenge

DARPA

**SHREDDER
CHALLENGE**

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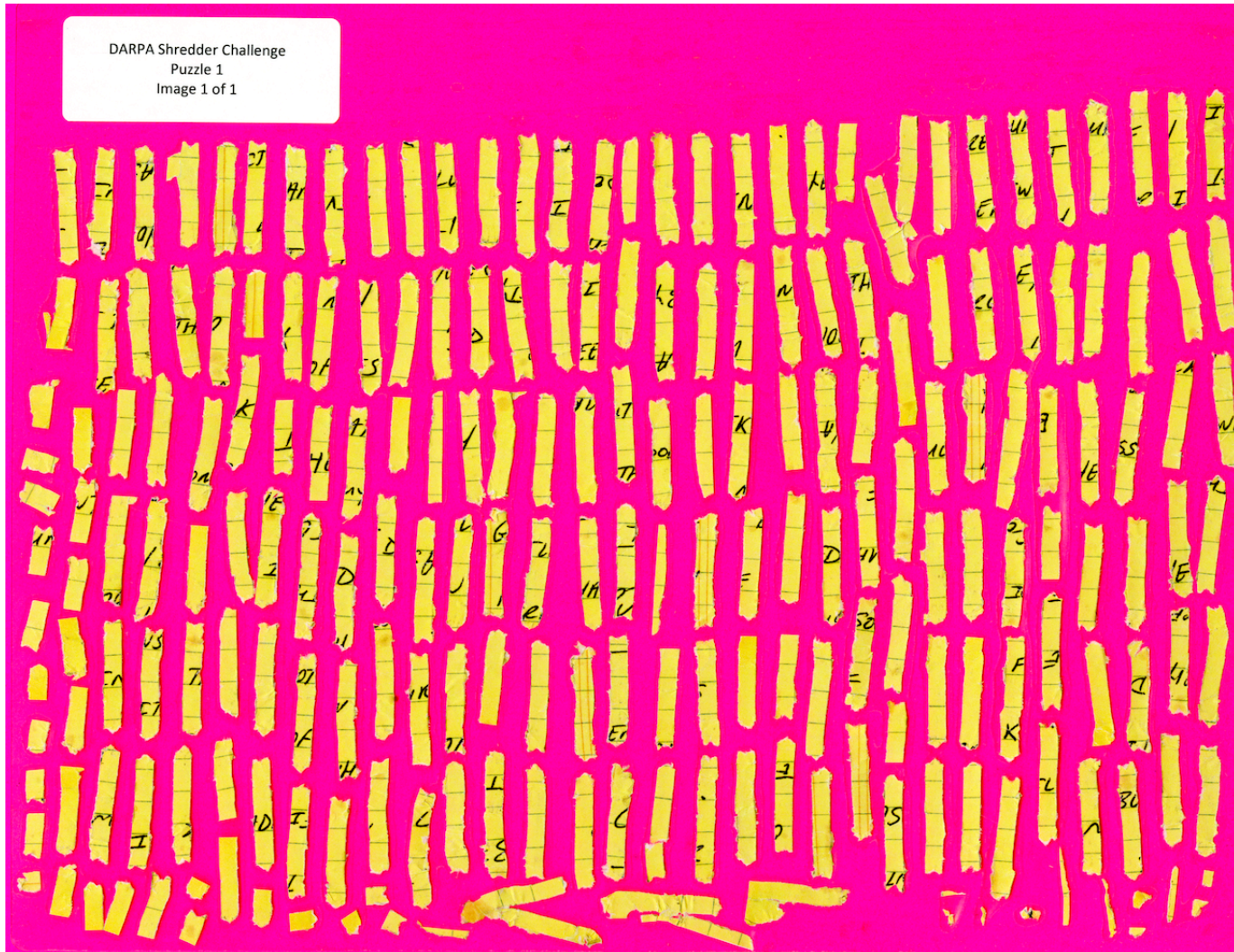


CONGRATULATIONS to "All Your Shreds Are Belong To U.S."!

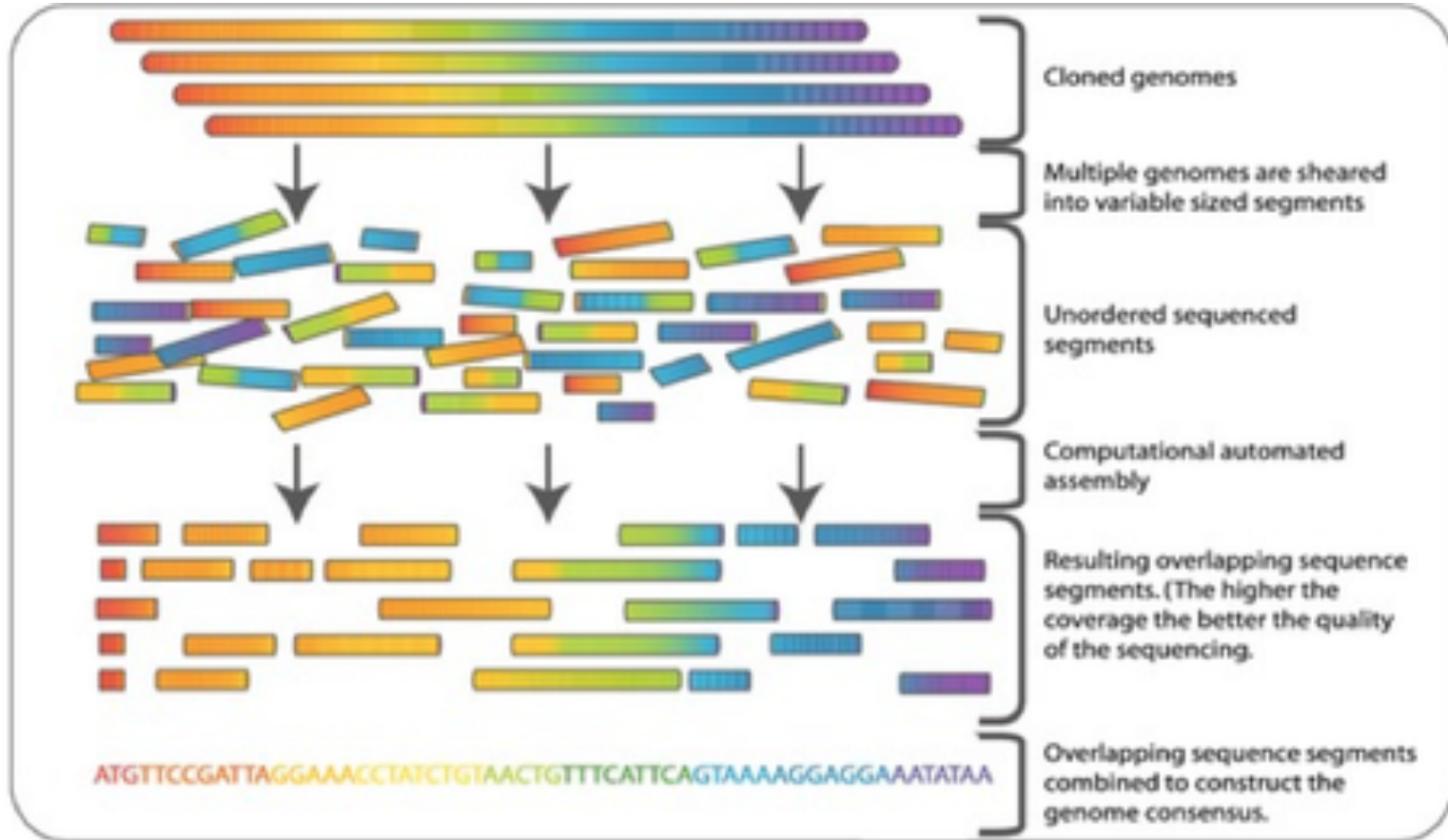
"All Your Shreds Are Belong To U.S." successfully reconstructed and solved all 5 Puzzles earning \$50,000!

[View the announcement](#)

DARPA Shredder Challenge

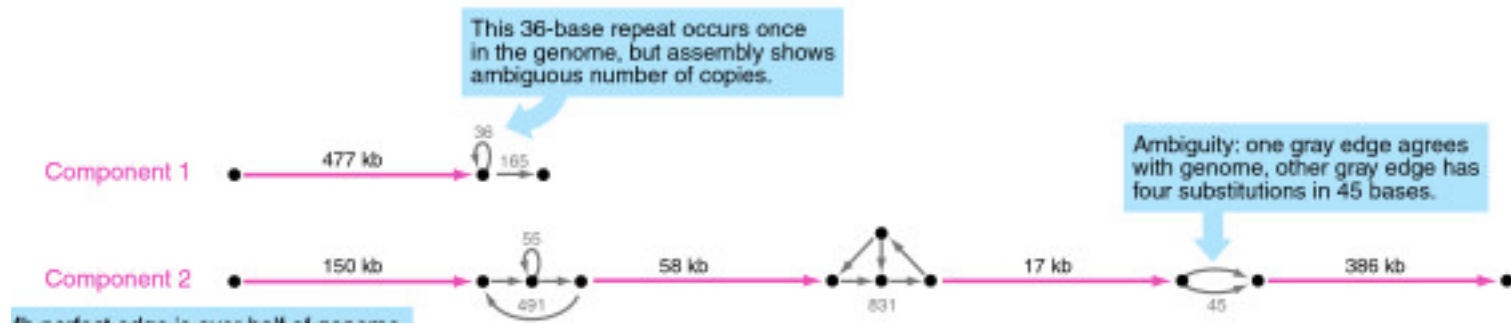


Shotgun Sequencing



Caveats

- Errors in reading
- Non-trivial assembly task: repeats in the genome

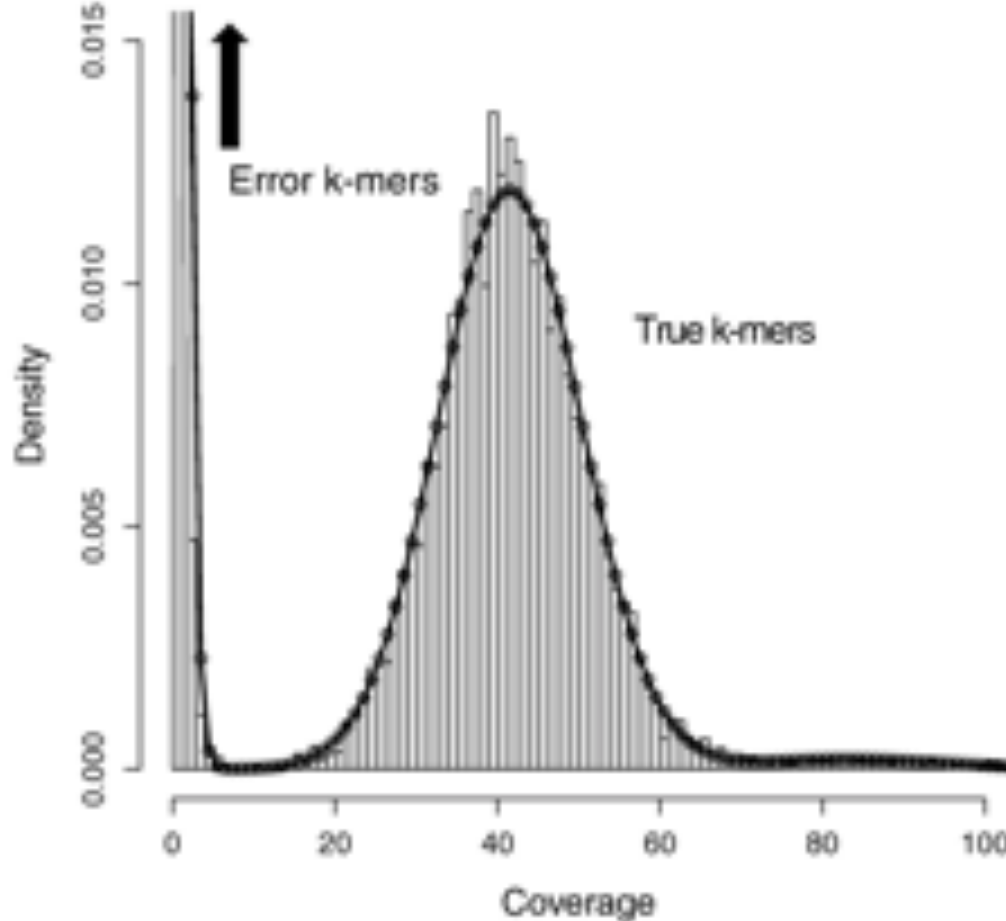


MacCallum et al., GB 2009

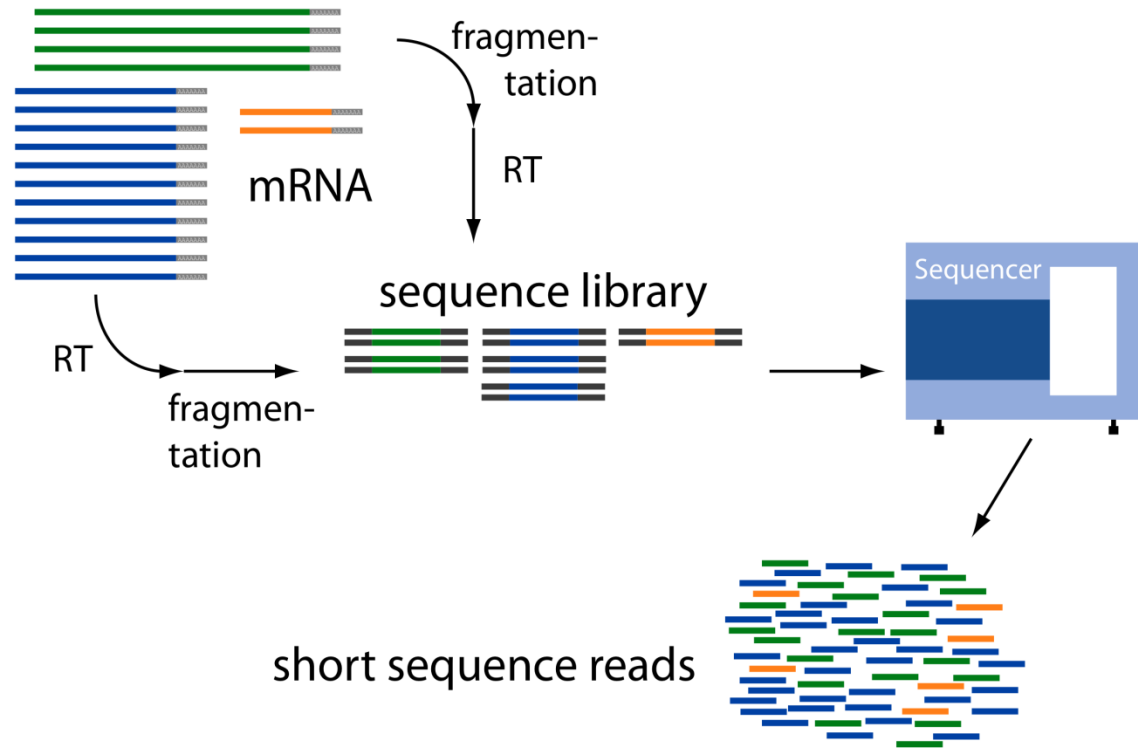
Error Correction in DNA sequencing

- The fragmentation happens at random locations of the molecules.
We expect all positions in the genome to have the same # number of reads

K-mers = substrings of length K of the reads. Errors create error k-mers.



Transcriptome Shotgun Sequencing (RNA-Seq)



Sequencing RNA transcripts.

Reminder:

- (mRNA) Transcripts are “expression products” of genes.
- Different genes having different expression levels so some transcripts are more or less abundant than others.

@Friedrich Miescher Laboratory

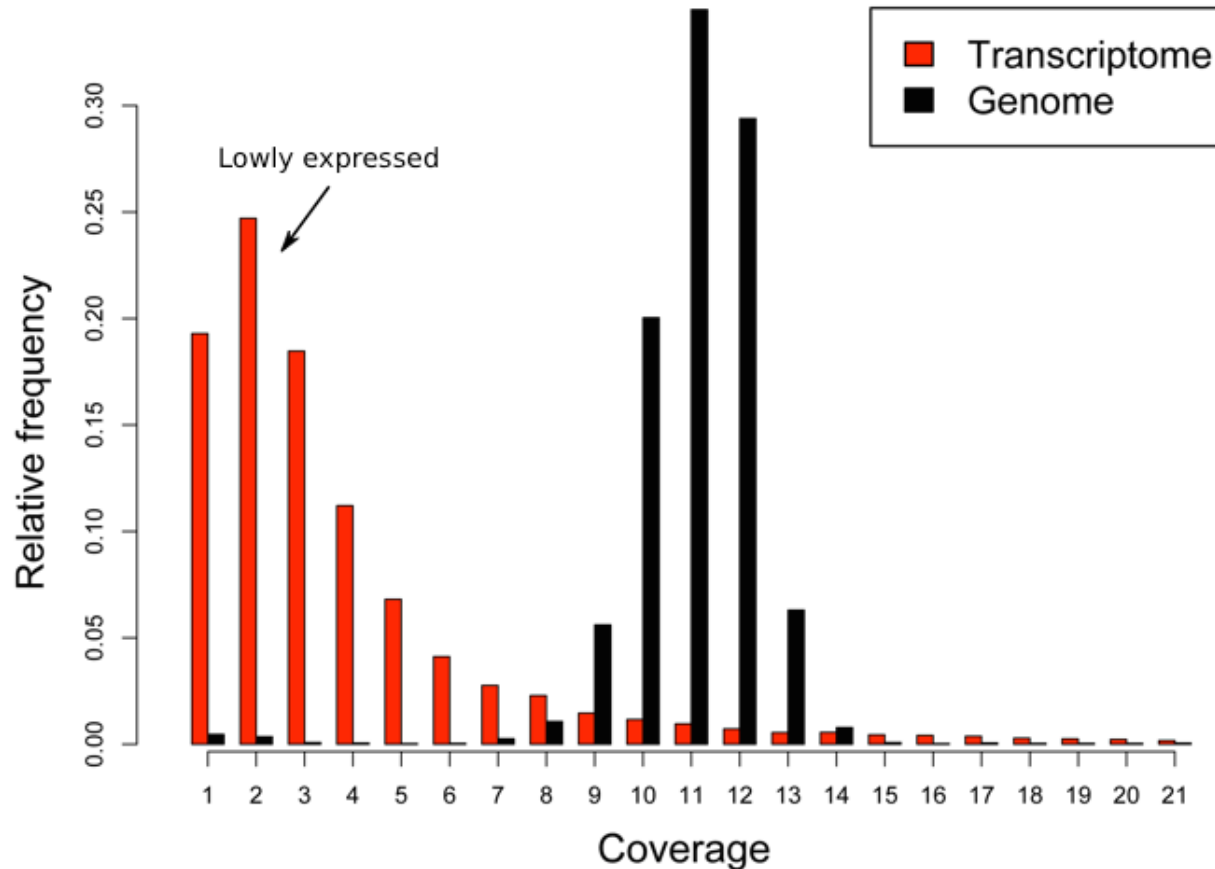
Challenges

- Large datasets: 10-100 millions reads of 75-150 bps.
- Memory efficiency: Too time consuming to perform out-memory processing of data.

DNA Sequencing + **others** : alternative slicing, RNA editing, post-transcription modification.

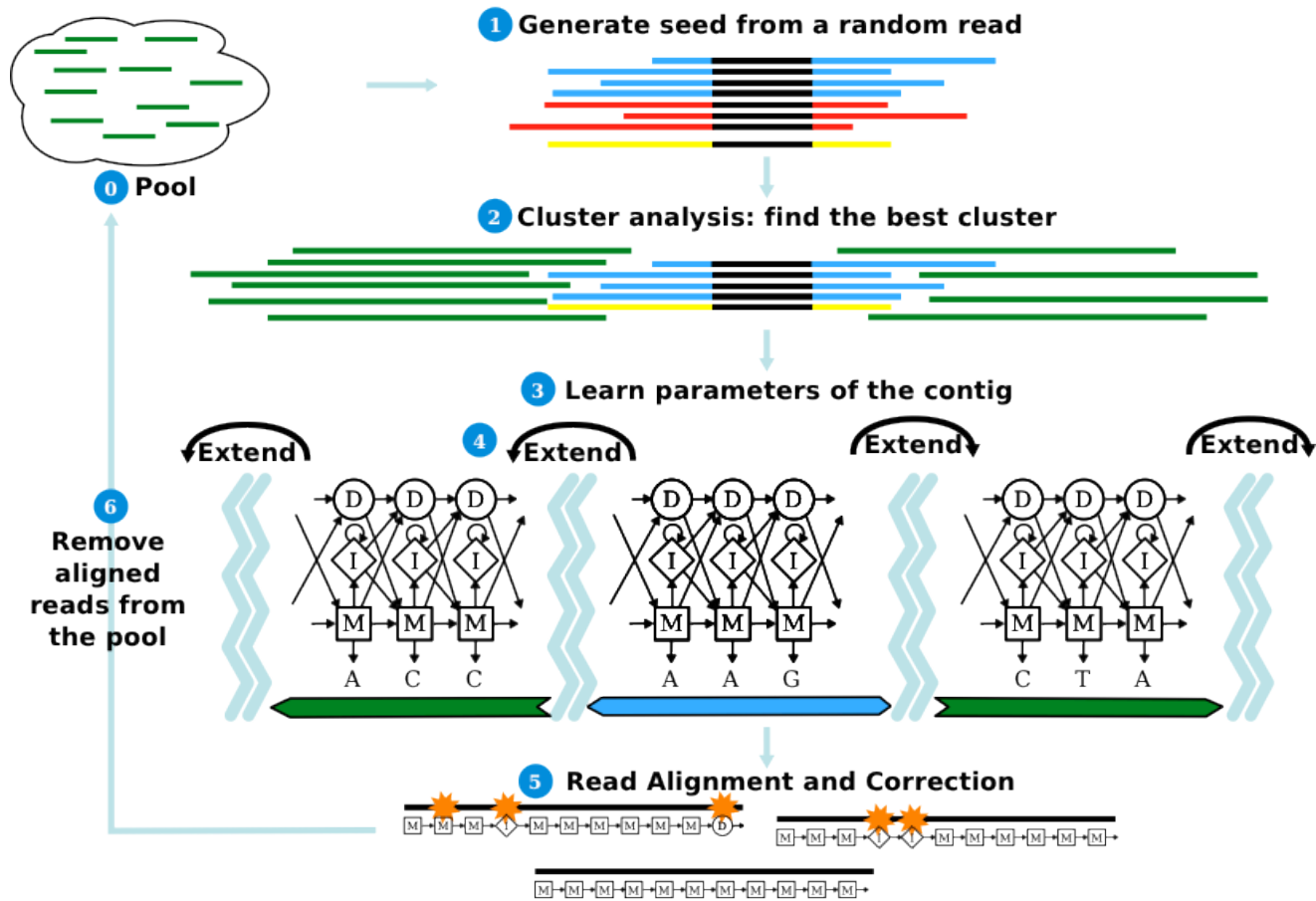
Errors are non uniformly distributed

- Some transcripts are more prone to errors
- Errors are harder to correct in reads from lowly expressed transcripts



SEECER

Error Correction + Consensus sequence estimation for RNA-Seq data



Key idea: HMM model

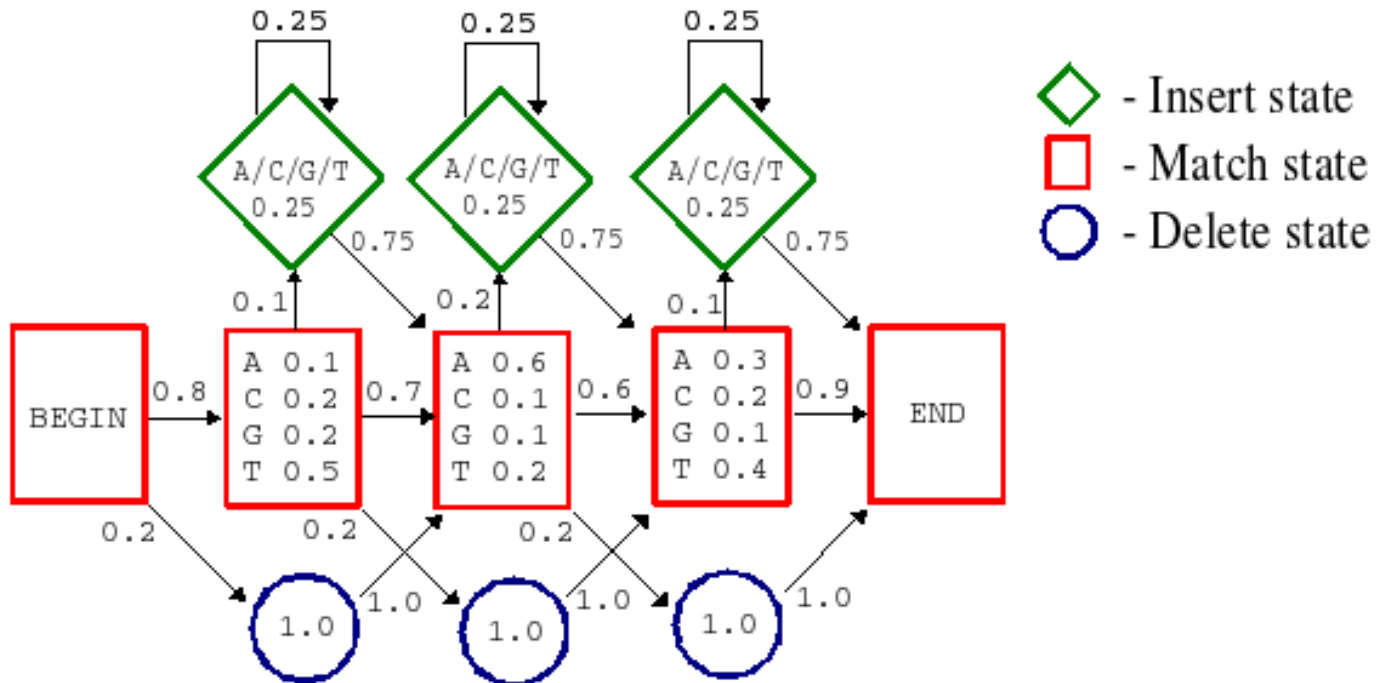
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Consensus	G	T	C	A	G	A	A	-	G	T	G	A	G	C	G	T	G	G	C	A	T	T	A	A	C	C	C	T	T	G	A	T	A	C	C	A	C	C	G	G	T	T	C	A	A	C	C										
Read 1	G	T	C	A	G	A	A	-	G	T	G	A	G	C	G	T	G	G	C	A	T	T	A	A	C	C	C	T	T	G	A	T	A	A	C	C	A	C	C	G	G	T	T	C	A	A	C	C									
	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40	40							
Read 2																																																									
Read 3																																																									
Read 4																																																									
Read 5																																																									
Read 6																																																									

Salmela et al., Bioinformatics 2011

The way sequencers work:

- Read letter by letter sequentially
- Possible errors: Insertion , Deletion or Misread of a nucleotide

Column	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47												
Consensus	G	T	C	A	G	A	A	-	G	T	G	A	G	C	G	T	G	G	C	A	T	T	A	A	C	C	C	T	T	G	A	T	A	C	C	A	C	C	G	G	T	T	C	A	A	C	C												
Read 1	G	T	C	A	G	A	A	-	G	T	G	A	G	C	G	T	G	G	C	A	T	T	A	A	C	C	C	T	T	G	A	T	A	A	C	C	C	T	T	G	A	T	A	A															
Read 2									A																																																		
Read 3	C	A	G	A	A	-	G	T	G	A	G	C	G	T	G	G	C	A	T	T	A	A	C	C	C	T	T	G	A	T	A	C	C	A	C	C	G	G	T	T	C	A	A																
Read 4																																																											
Read 5																																																											
Read 6																																																											



Building (Learning) the HMMs and Making Corrections (Inference)

Learning = Expectation-Maximization

Inference = Viterbi algorithm

Seeding:

Guessing possible reads using k-mer overlaps.

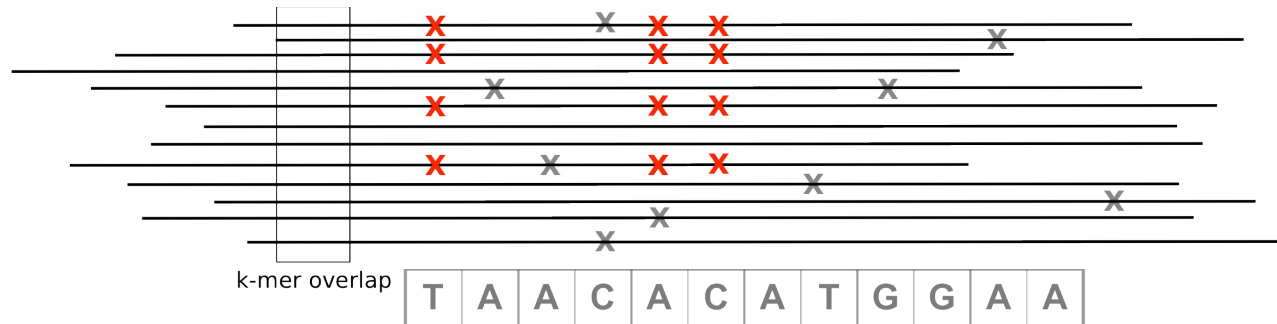
Constructing the HMM from these reads.

Speed up:

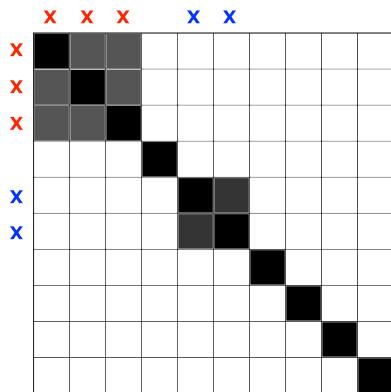
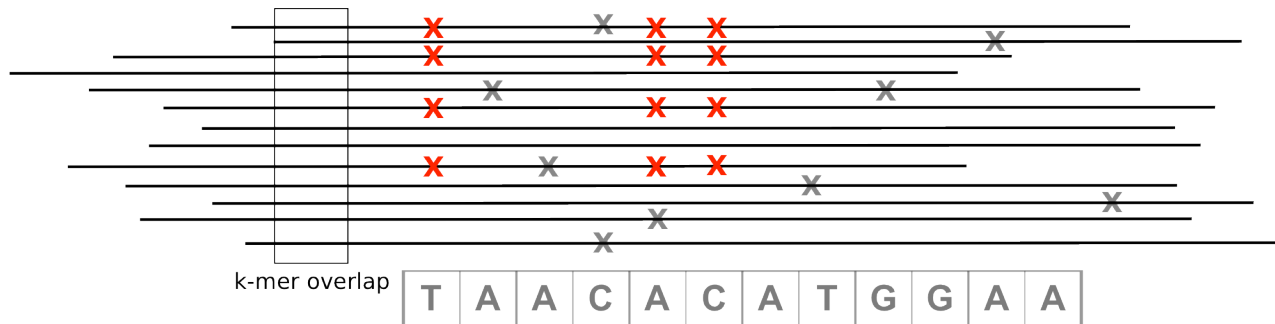
The k-mer overlaps yield approximate multiple alignments of reads.

We can learn HMM parameters from this directly.

Clustering to improve seeding

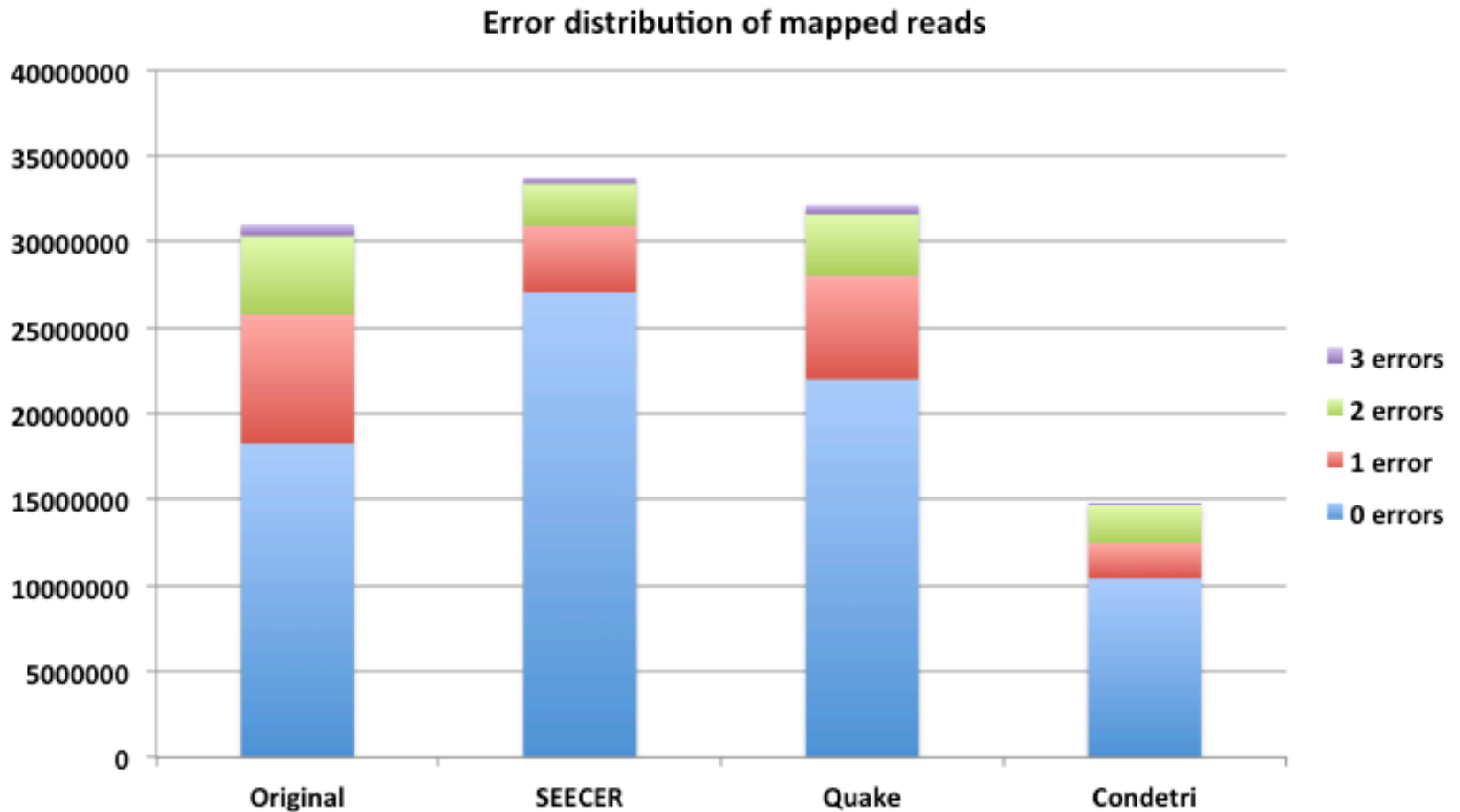


Real biological differences should be supported by a set of reads with similar mismatches to the consensus

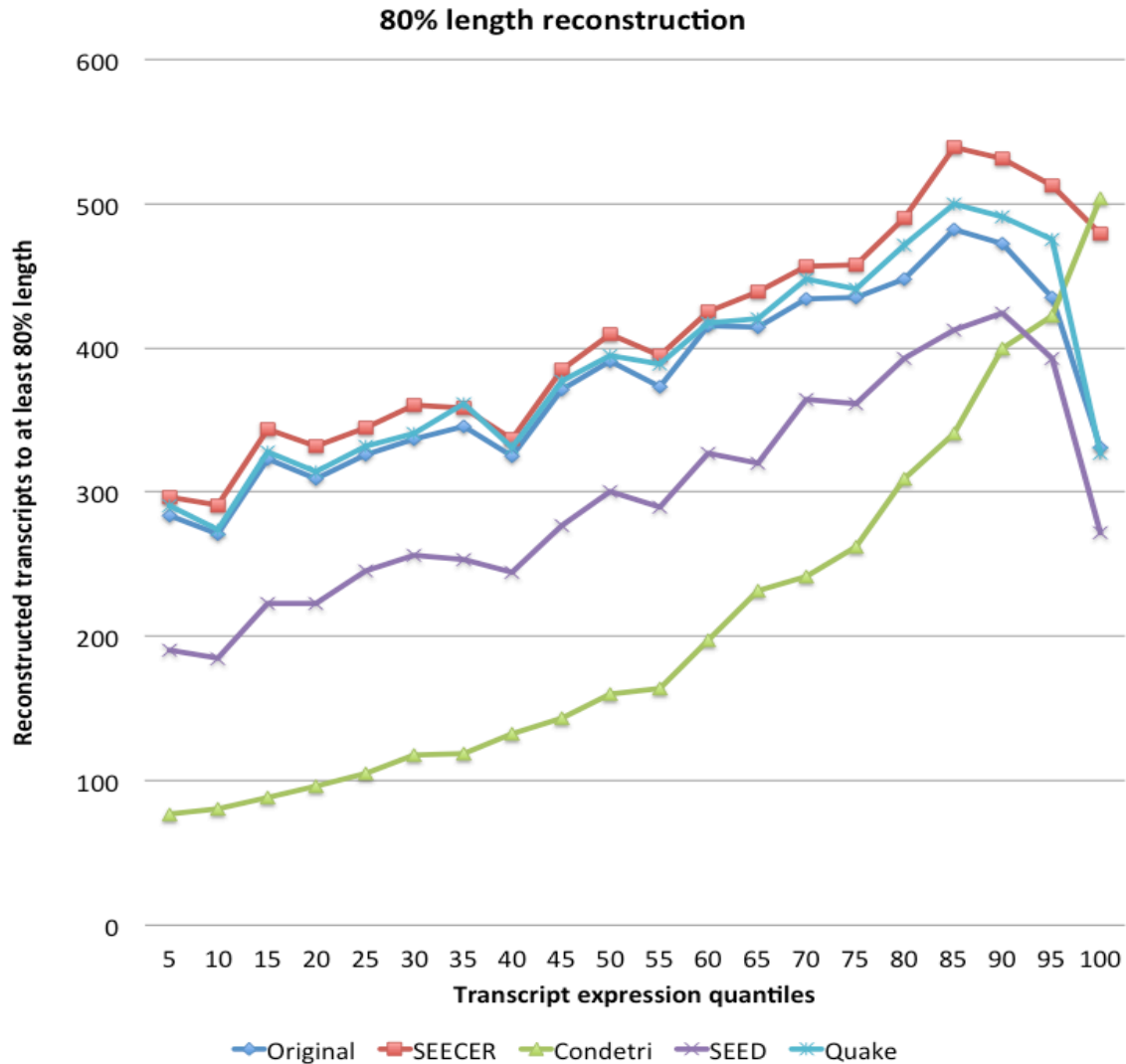


1. Clustering positions with mismatches to identify clusters of correlated positions.
2. Build a similarity matrix between these positions.
3. Use Spectral clustering to find clusters of correlated positions.
4. Filter reads have mismatches in these clusters.

Comparison to other methods



Using the corrected reads, the assembler can recover **MORE** transcripts

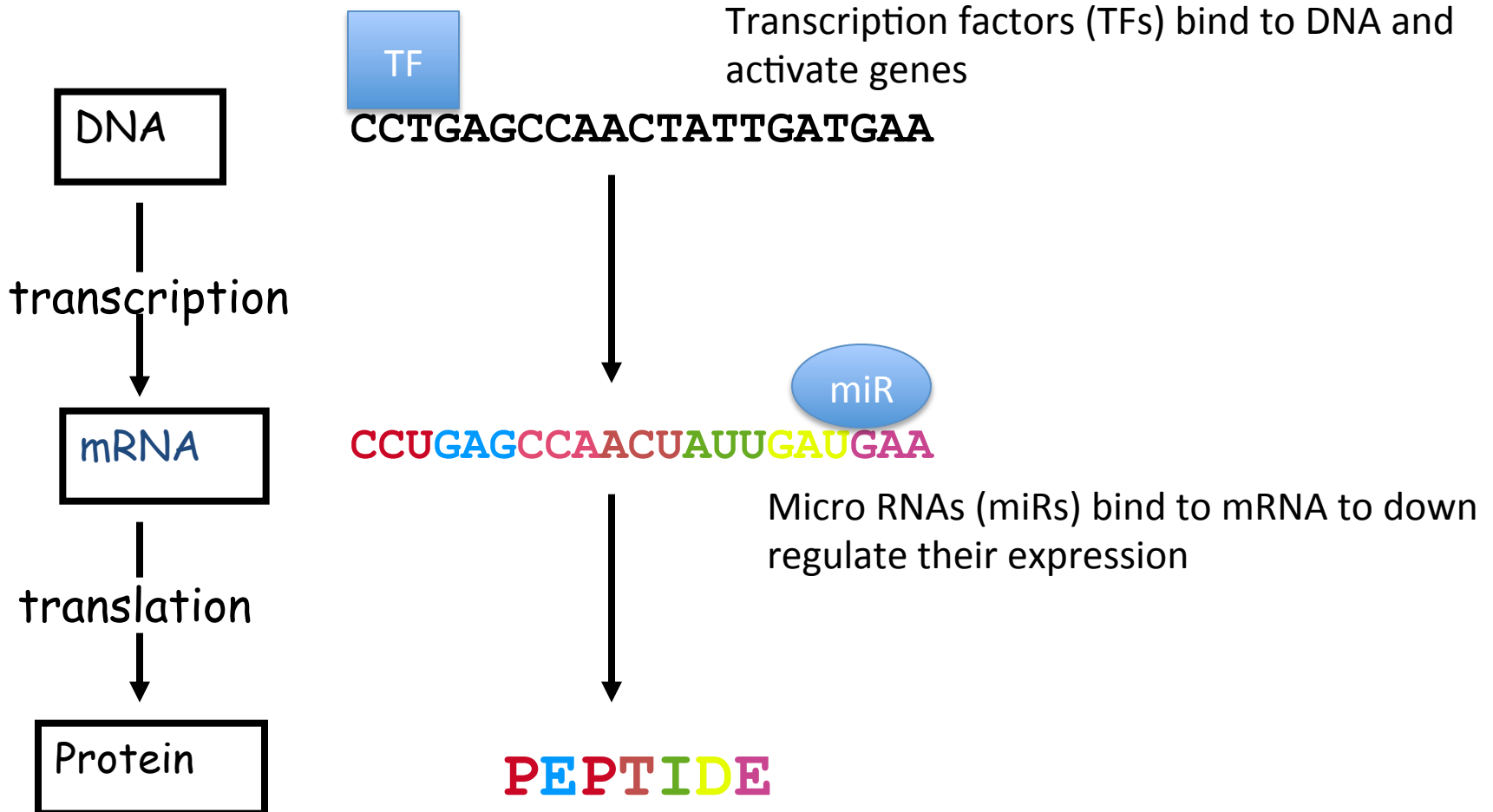


Things that work

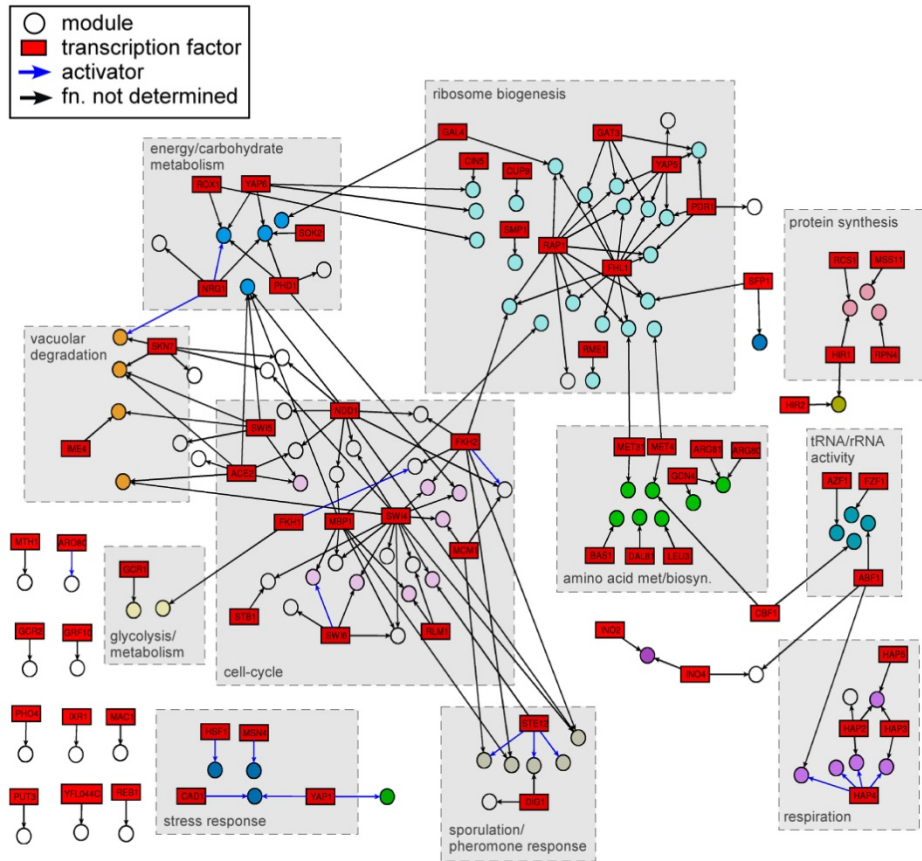
- Approximate learning to speed up on large datasets.
- In real world, one technique is not enough. A solution involves using many techniques.
- Precision and Recall are trade-offs.

Central dogma

Different regulators control the information flow from DNA to protein



Integrating expression and protein-DNA interaction data

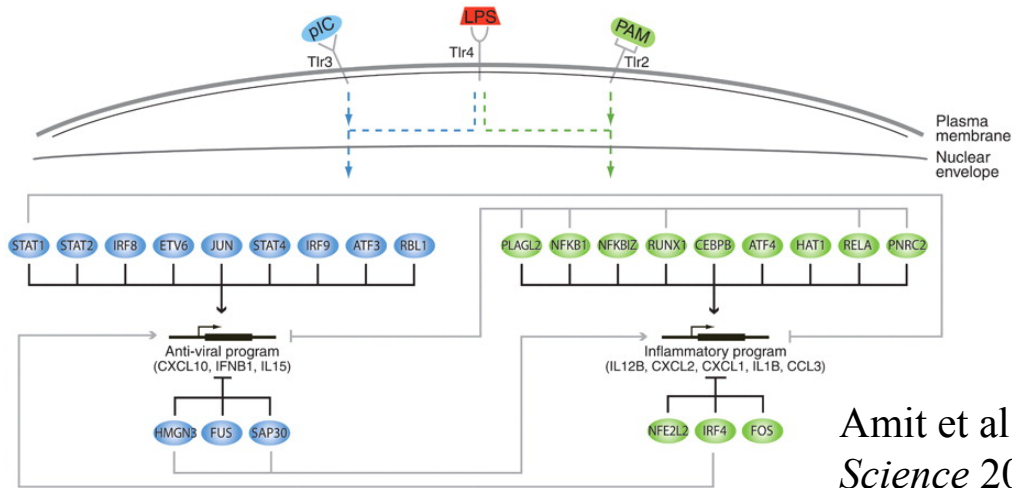


Lee *et al* Science 2002

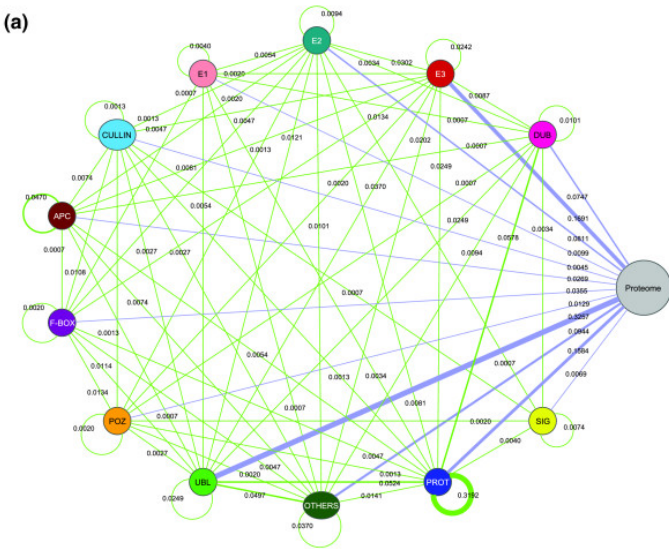
Bar-Joseph *et al* Nature Biotechnology 2003

module categories			
tRNA/rRNA activity	respiration	carbohydrate metabolism	vacuolar degradation
ribosome biogenesis	stress response	sporulation/pheromone response	cell cycle
amino acid met./biosyn.	protein synthesis	chromosome/histone	unknown
glycolysis/metabolism	fermentation	lipid/fatty acid biosyn.	

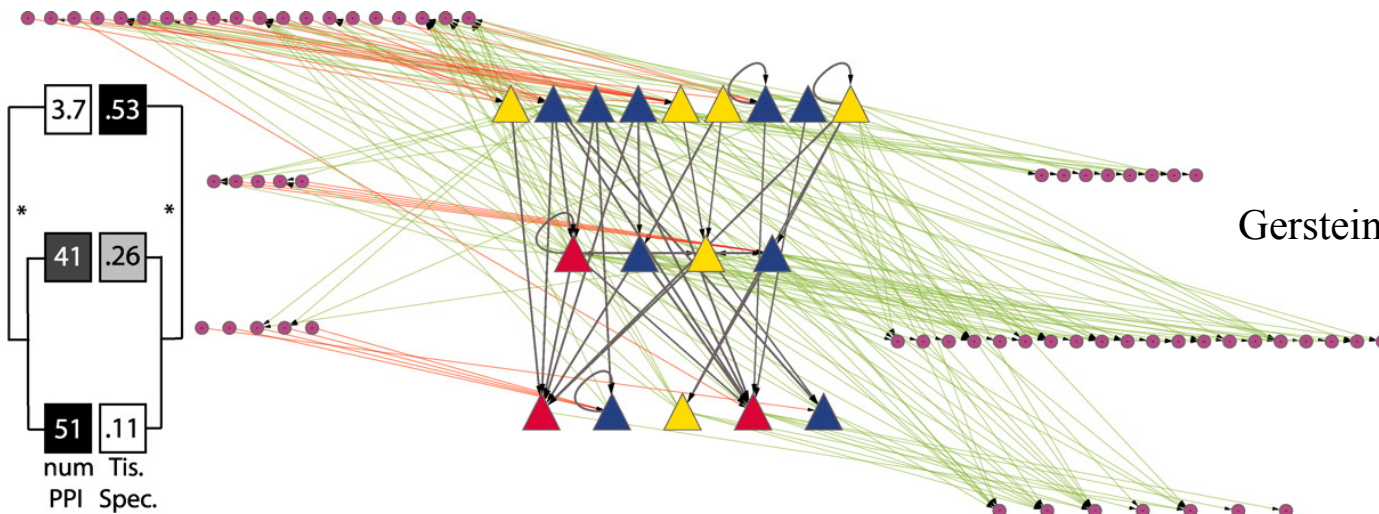
Methods for reconstructing networks in cells (a)



Amit et al
Science 2009



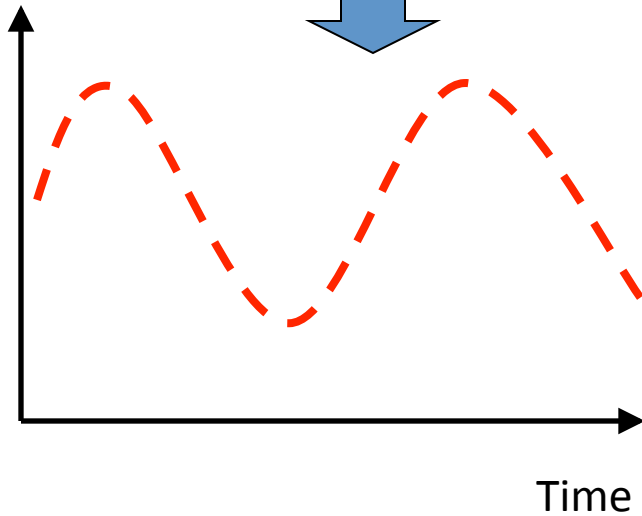
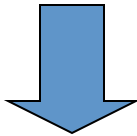
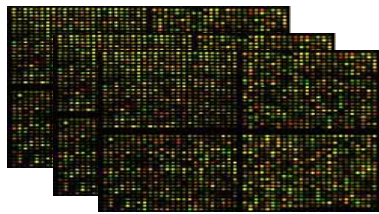
Venancio et al *Genome Biology* 2009



Gerstein et al *Science* 2010

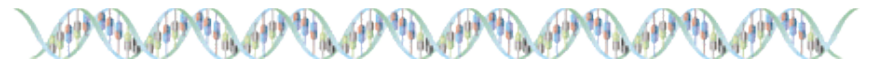
Key problem: Most high-throughput data is static

Time-series measurements



Static data sources

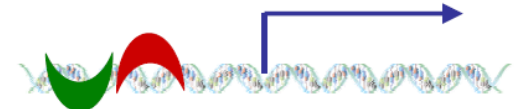
Sequencing



motif



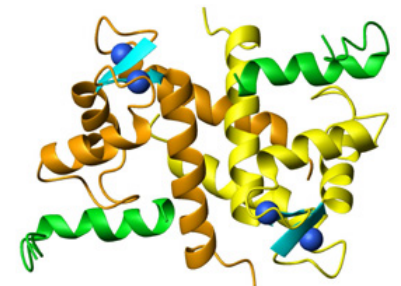
CHIP-chip



microarray

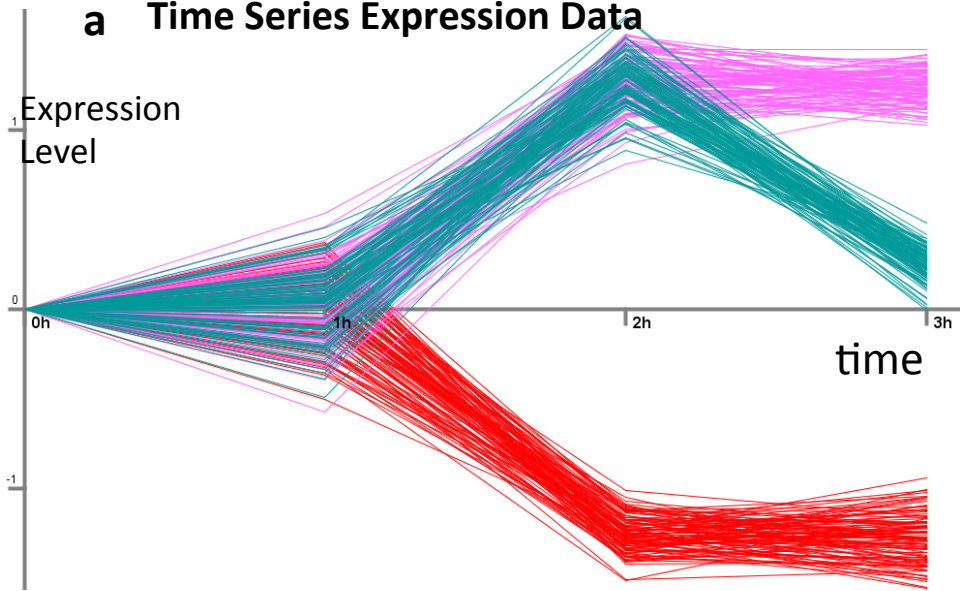


PPI

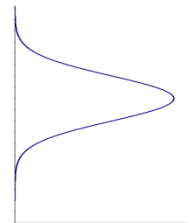
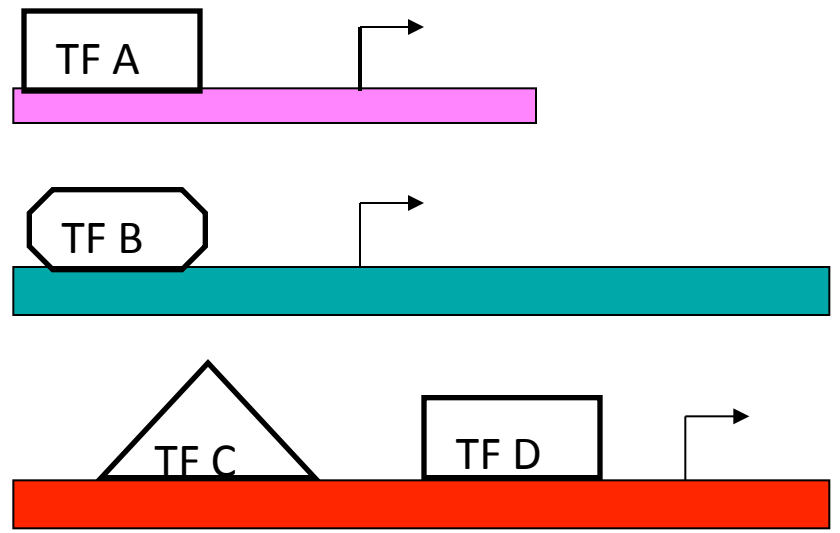


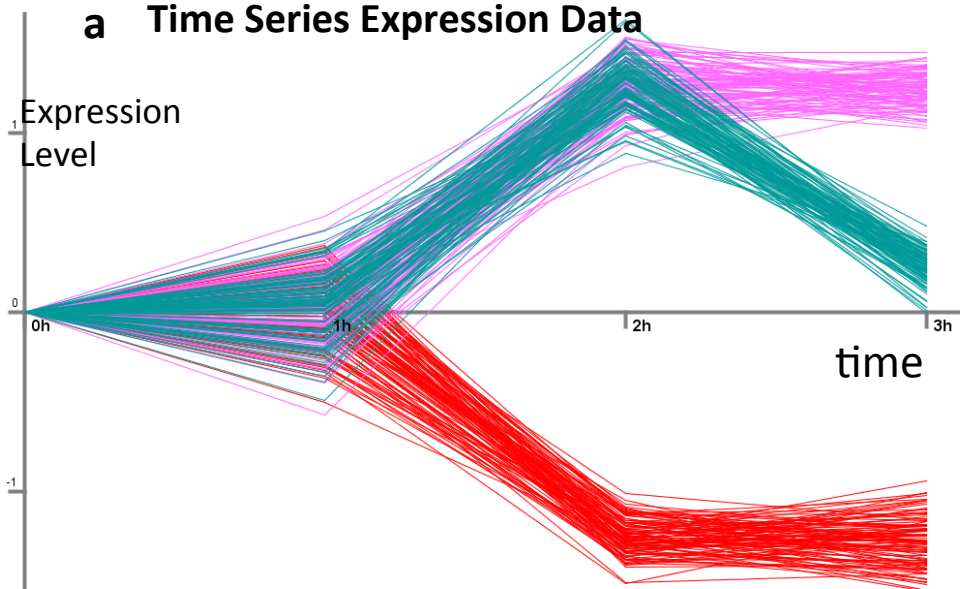
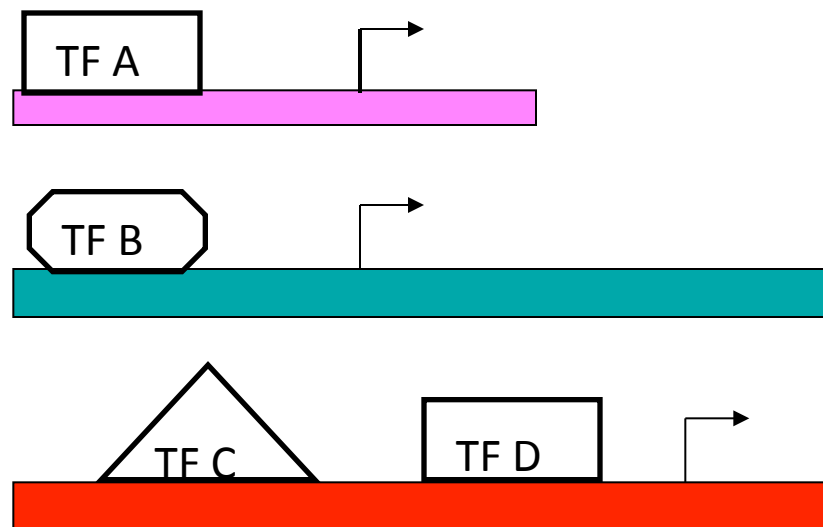
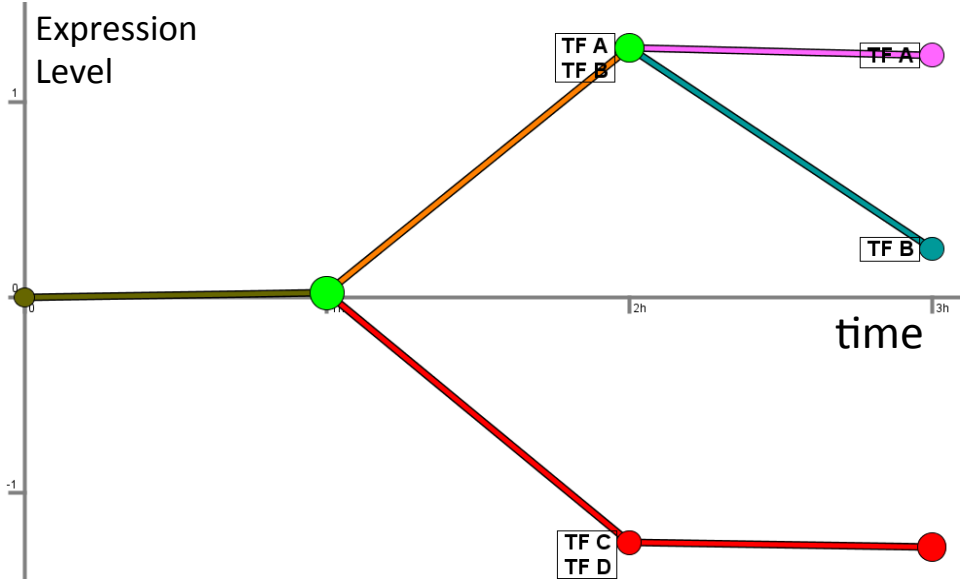
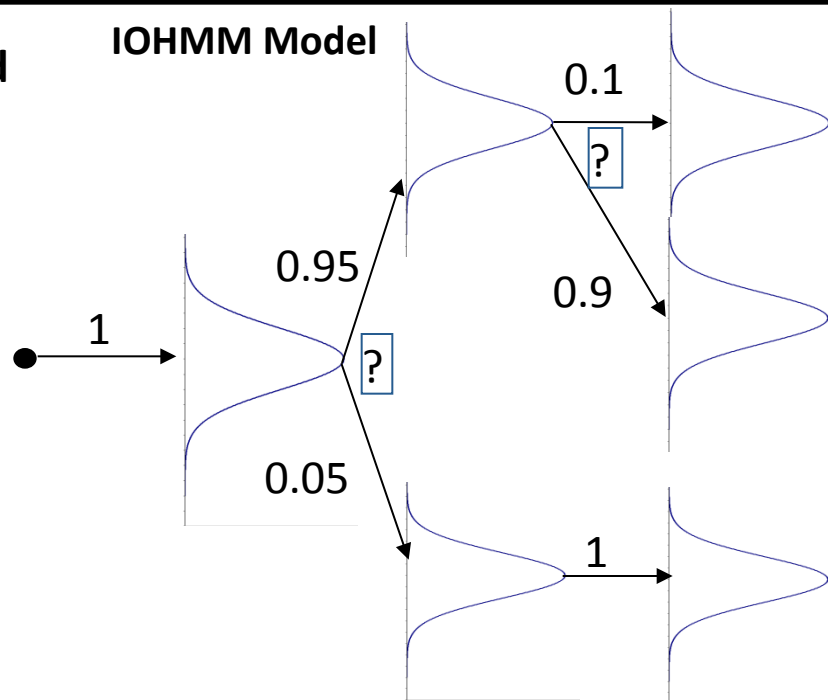
DREM: Dynamic Regulatory Events Miner

a Time Series Expression Data

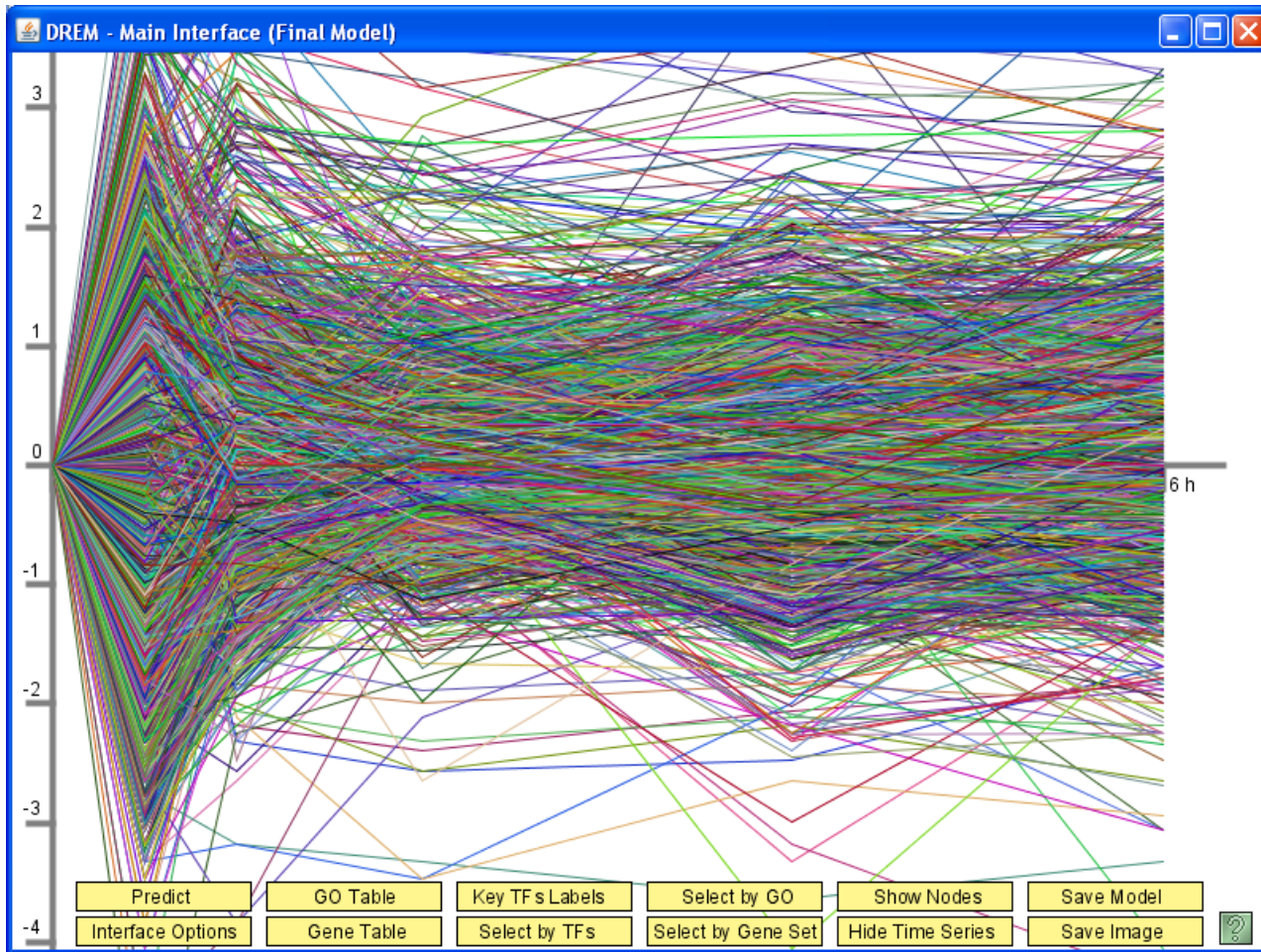


b Static TF-DNA Binding Data

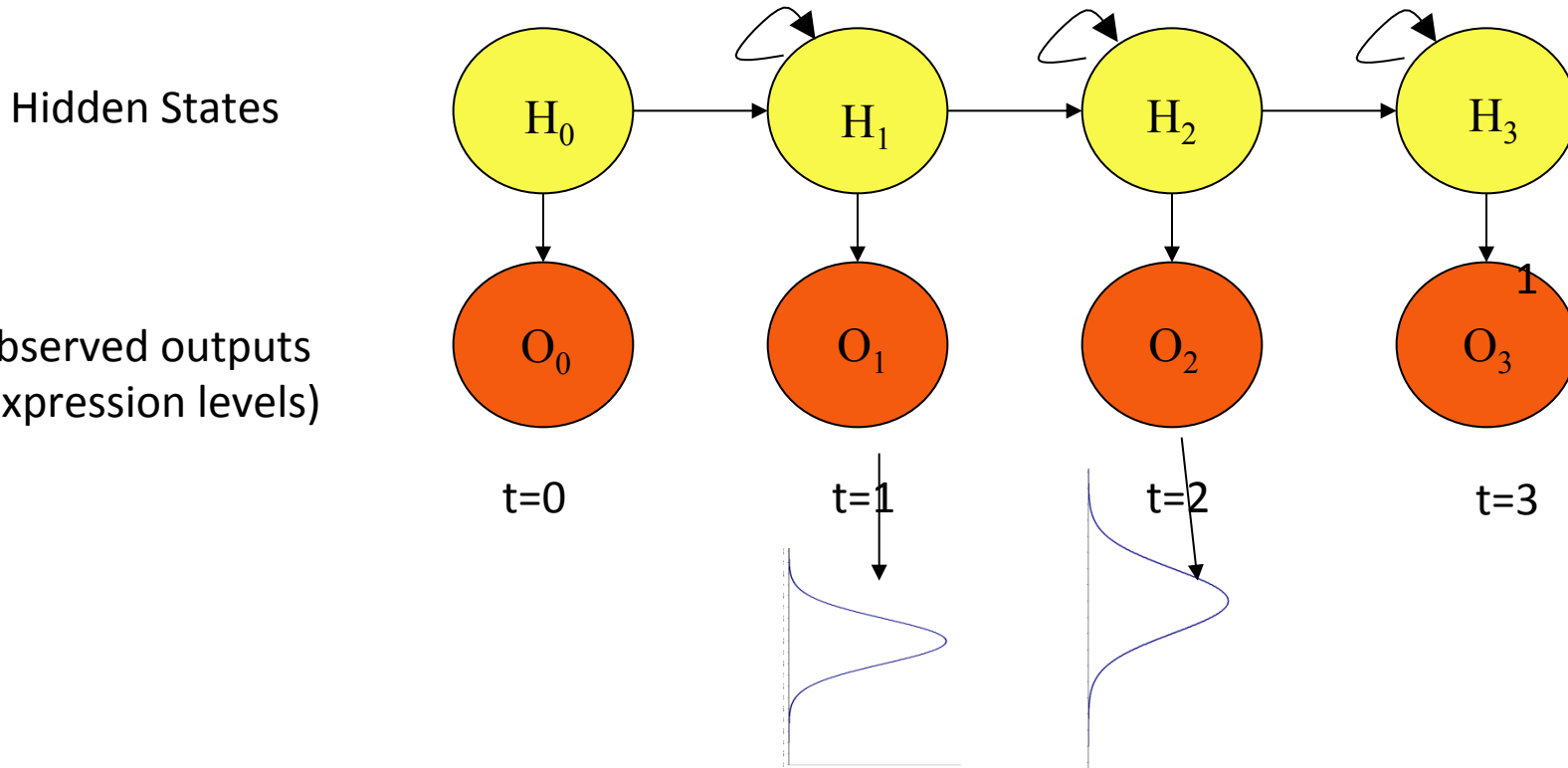


a Time Series Expression Data**b Static TF-DNA Binding Data****c Model Structure****d IOHMM Model**

Things are a bit more complicated: Real data



A Hidden Markov Model



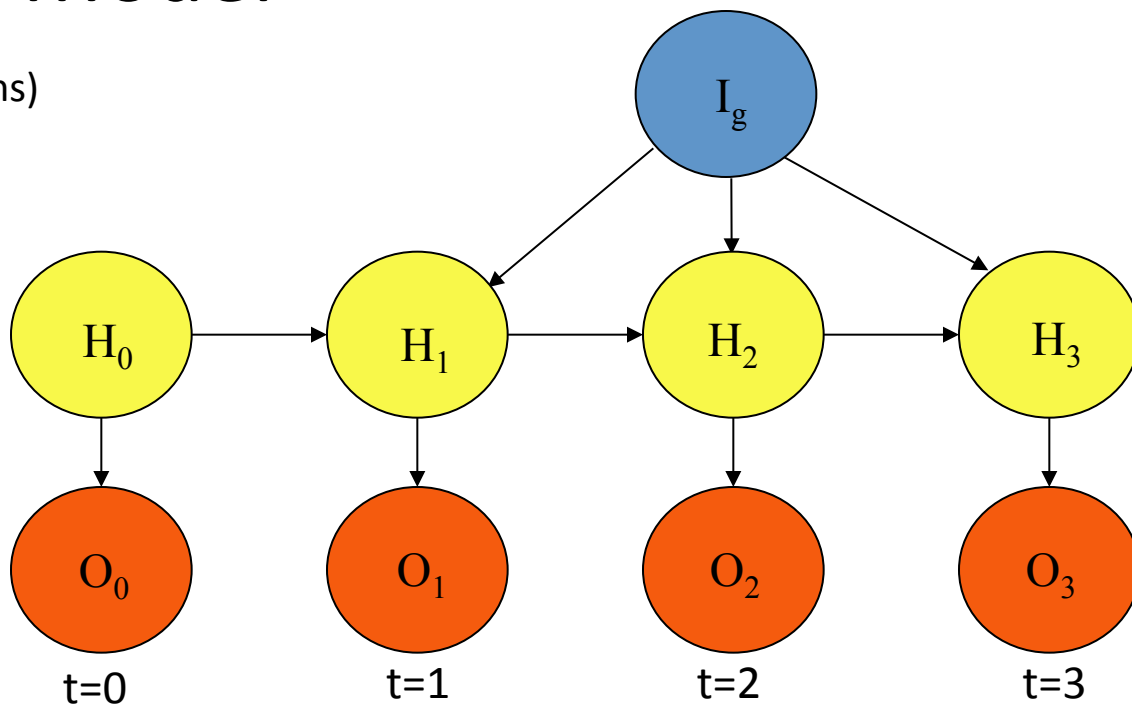
$$L(H, O; \Theta) = \prod_{i=1}^n \left[\prod_{t=1}^T p(O_t(i) | H_t(i)) \right] \left[\prod_{t=2}^T p(H_t(i) | H_{t-1}(i)) \right]$$

Input – Output Hidden Markov Model

Input (Static TF-gene interactions)

Hidden States (transitions between states form a tree structure)

Emissions (Distribution of expression values)



Log Likelihood

But how do we express these conditional probabilities?

$$r(G|M) = \sum_{g \in G} \log \sum_{q \in Q} \prod_{t=1}^{n-1} \underbrace{f_{q(t)}(o_g(t))}_{\text{Product over all Gaussian emission density values on path}} \prod_{t=1}^{n-1} \underbrace{P(H_t = q(t) | H_{t-1} = q(t-1), I_g)}_{\text{Product over all transition probabilities on path}}$$

Sum over all genes

Sum over all paths Q

Input-Output Hidden Markov Model

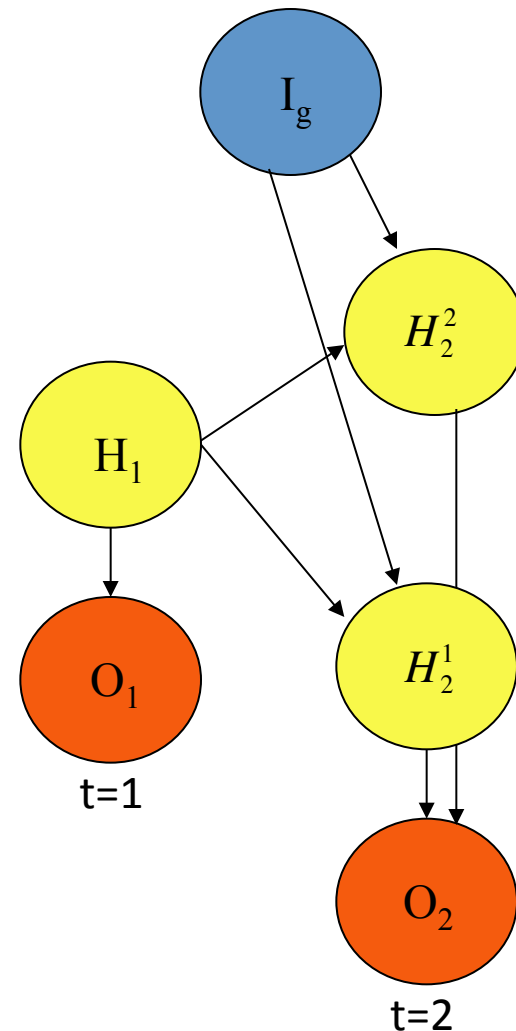
learning the transition probabilities

How do compute P for a state with 2 children?

We can write it as a logistic regression classification problem!

$$P(H_2^1 = q(2) | H_1 = q(1), I_g) = ?$$

$$P(H_2^2 = q(2) | H_1 = q(1), I_g) = ?$$



Input-Output Hidden Markov Model

learning the transition probabilities

How do compute P for a state with 2 children?

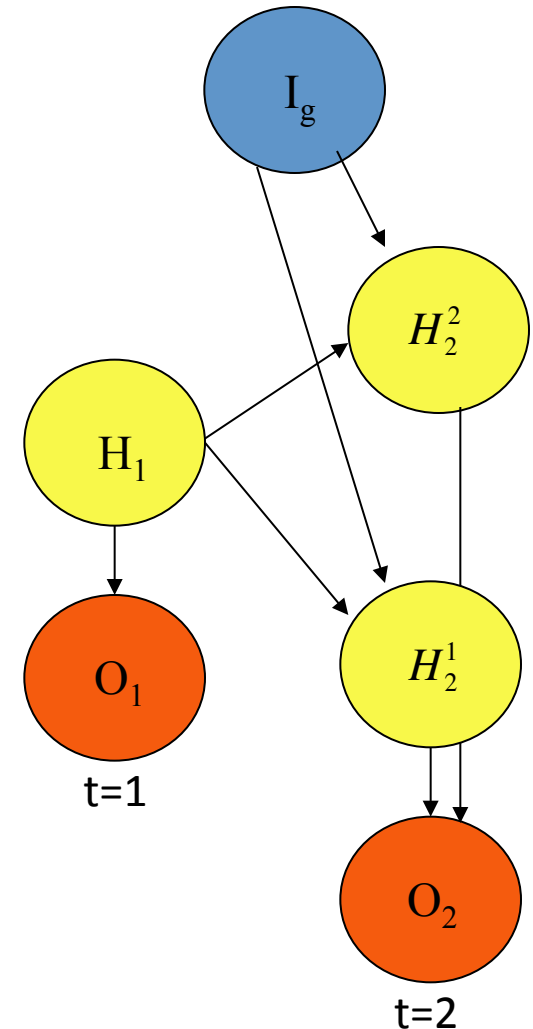
We can write it as a logistic regression classification problem!

$$P(H_2^1 = q(2) | H_1 = q(1), I_g) = \frac{1}{1 + \exp(w_0 + \sum_r I_{g,r} w_r)}$$

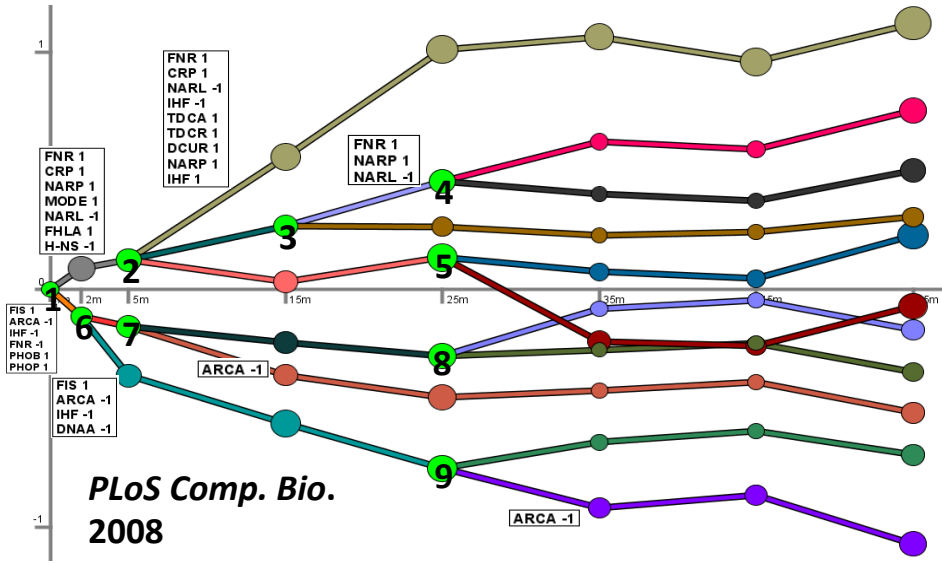
Sum over all regulators

Optimize w_r 's with a logistic regression classifier

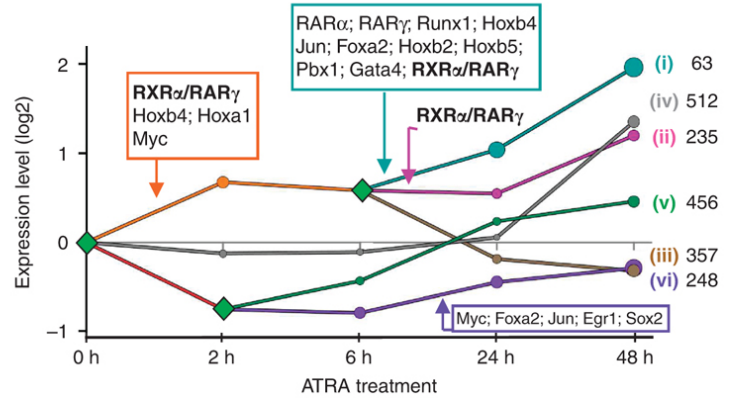
$$\hat{W} = \operatorname{argmax}_W \left[\underbrace{l(W)}_{\text{likelihood with parameters } W} + \underbrace{\ln p(W)}_{L_1\text{-penalty to promote sparsity}} \right]$$



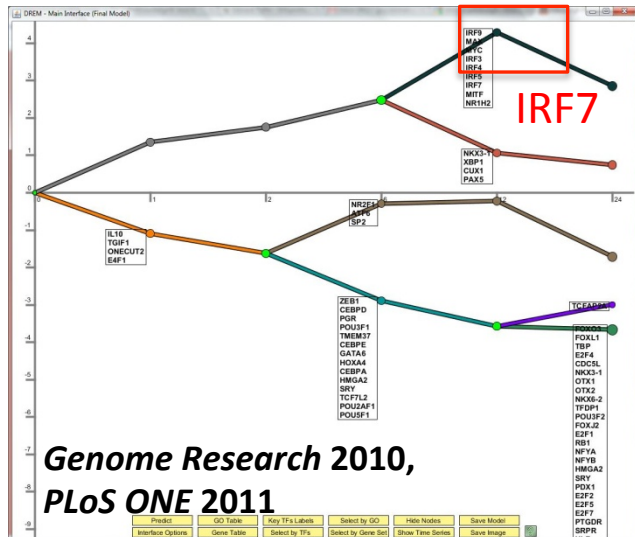
E. coli. response



Stem cells differentiation

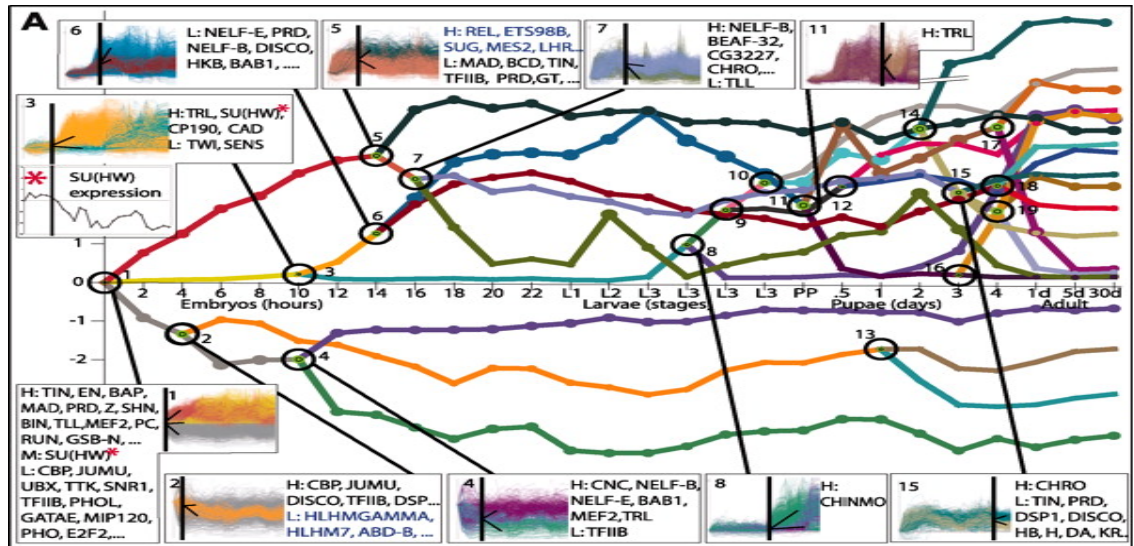


Mouse Immune response



Fly development

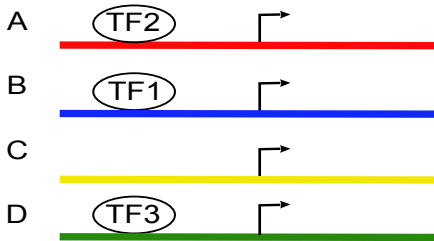
Science 2010



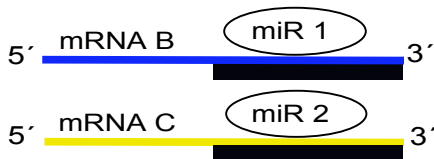
mirDREM

1. Regulatory interactions

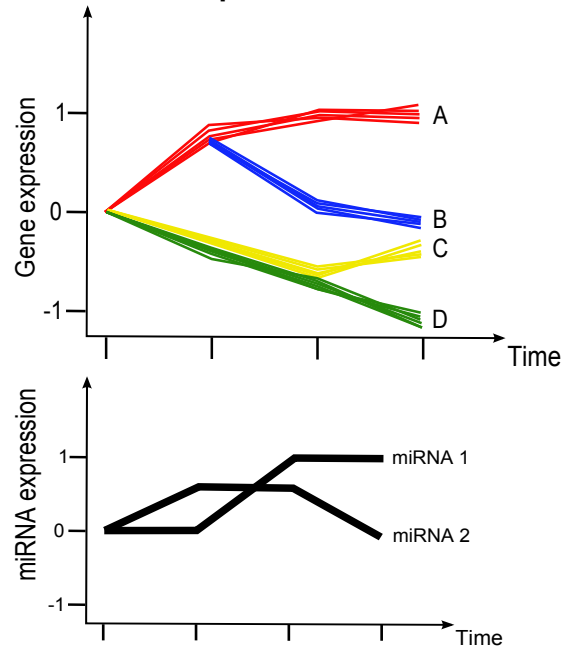
Transcriptional regulation



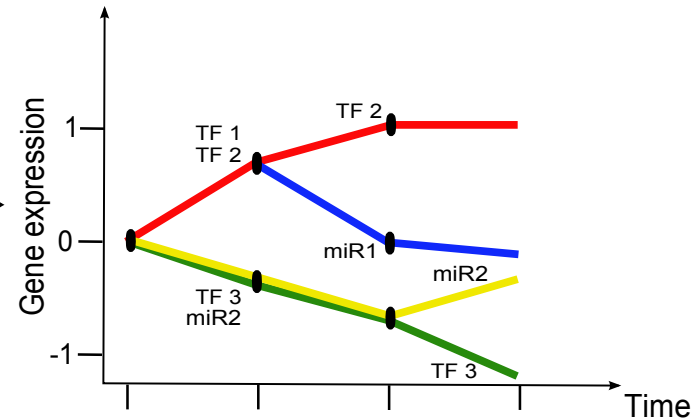
Post-transcriptional regulation



2. Expression data



3. Dynamic network



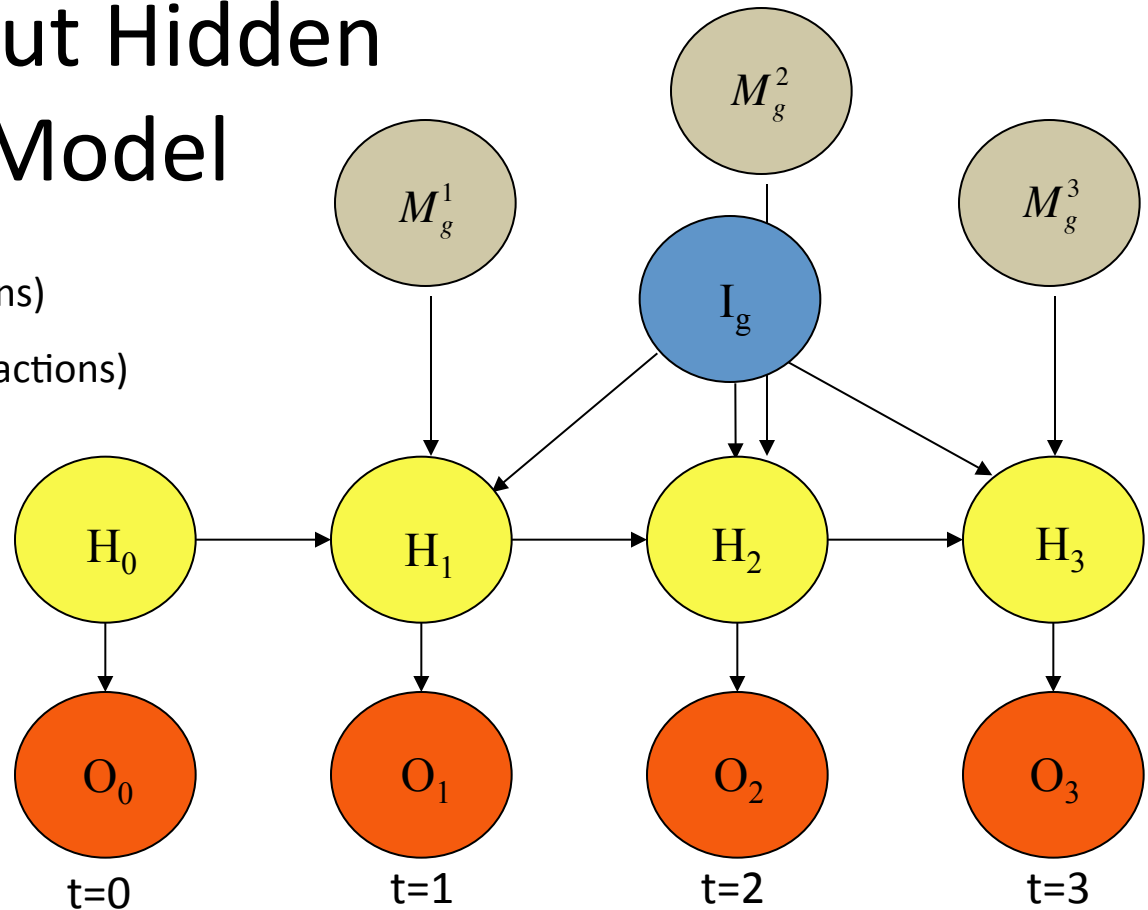
Input – Output Hidden Markov Model

Input (**Static** TF-gene interactions)

(**Dynamic** miR-gene interactions)

Hidden States (transitions between states form a tree structure)

Emissions (Distribution of expression values)



Log Likelihood

But how do we express these conditional probabilities?

$$r(G|M) = \sum_{\underbrace{g \in G}_{\text{Sum over all genes}}} \log \sum_{\underbrace{q \in Q}_{\text{Sum over all paths } Q}} \prod_{t=1}^{n-1} \underbrace{f_{q(t)}(o_g(t))}_{\text{Product over all Gaussian emission density values on path}} \prod_{t=1}^{n-1} \underbrace{P(H_t = q(t) | H_{t-1} = q(t-1), I_g, M_g^t)}_{\text{Product over all transition probabilities on path}}$$

Input-Output Hidden Markov Model

learning the transition probabilities

How do compute P for a state with 2 children?

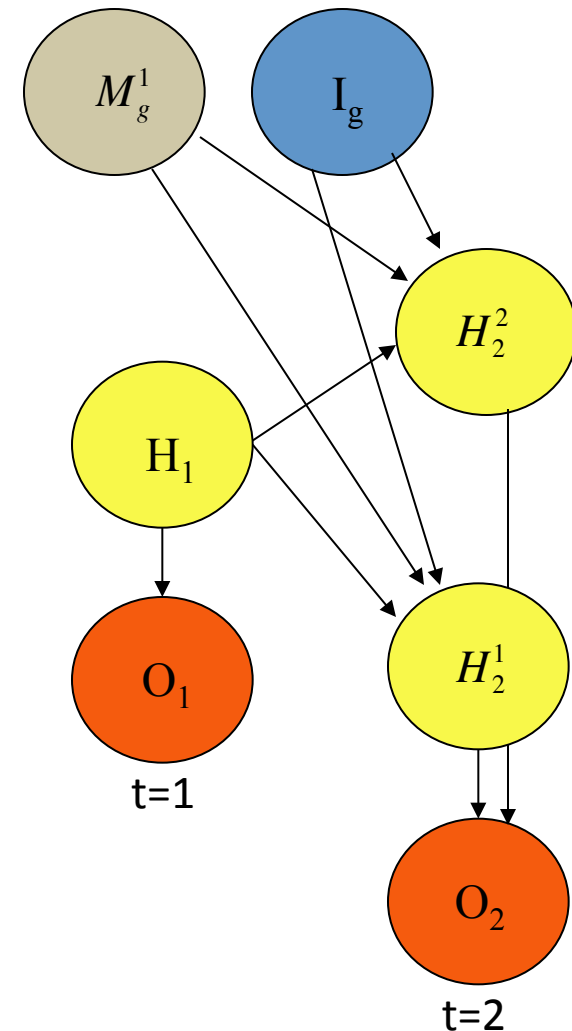
We can write it as a logistic regression classification problem!

$$P(H_2 = q(2) | H_1 = q(1), I_g) = \frac{1}{1 + \exp(w_0 + \underbrace{\sum_r I_{g,r} w_r}_{\text{Sum over all TFs}} + \underbrace{\sum_m M_{g,r}^1 w_m}_{\text{Sum over all miRs}})}$$

Optimize w_r 's with a **constrained** logistic regression classifier

$$\hat{W} = \underset{W}{\operatorname{argmax}} \left[\underbrace{l(W)}_{\text{likelihood with parameters } W} + \underbrace{\ln p(W)}_{\text{L}_1\text{-penalty to promote sparsity}} \right]$$

$$\text{s.t. } w_i \leq 0, i \in \text{miRNAs}$$



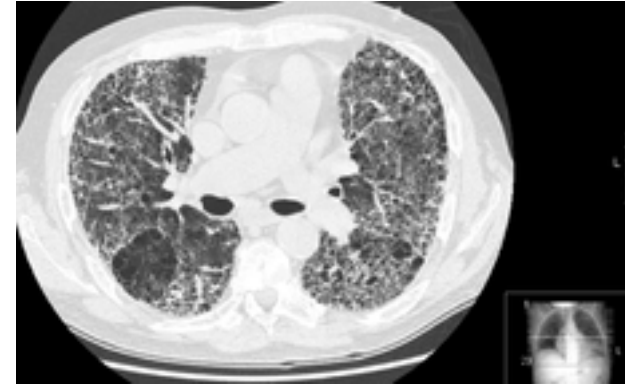
Application to mouse data to understand human lung disease

Idiopathic pulmonary fibrosis (IPF)

-100,000 people are affected (USA)

-about 30,000 new cases each year

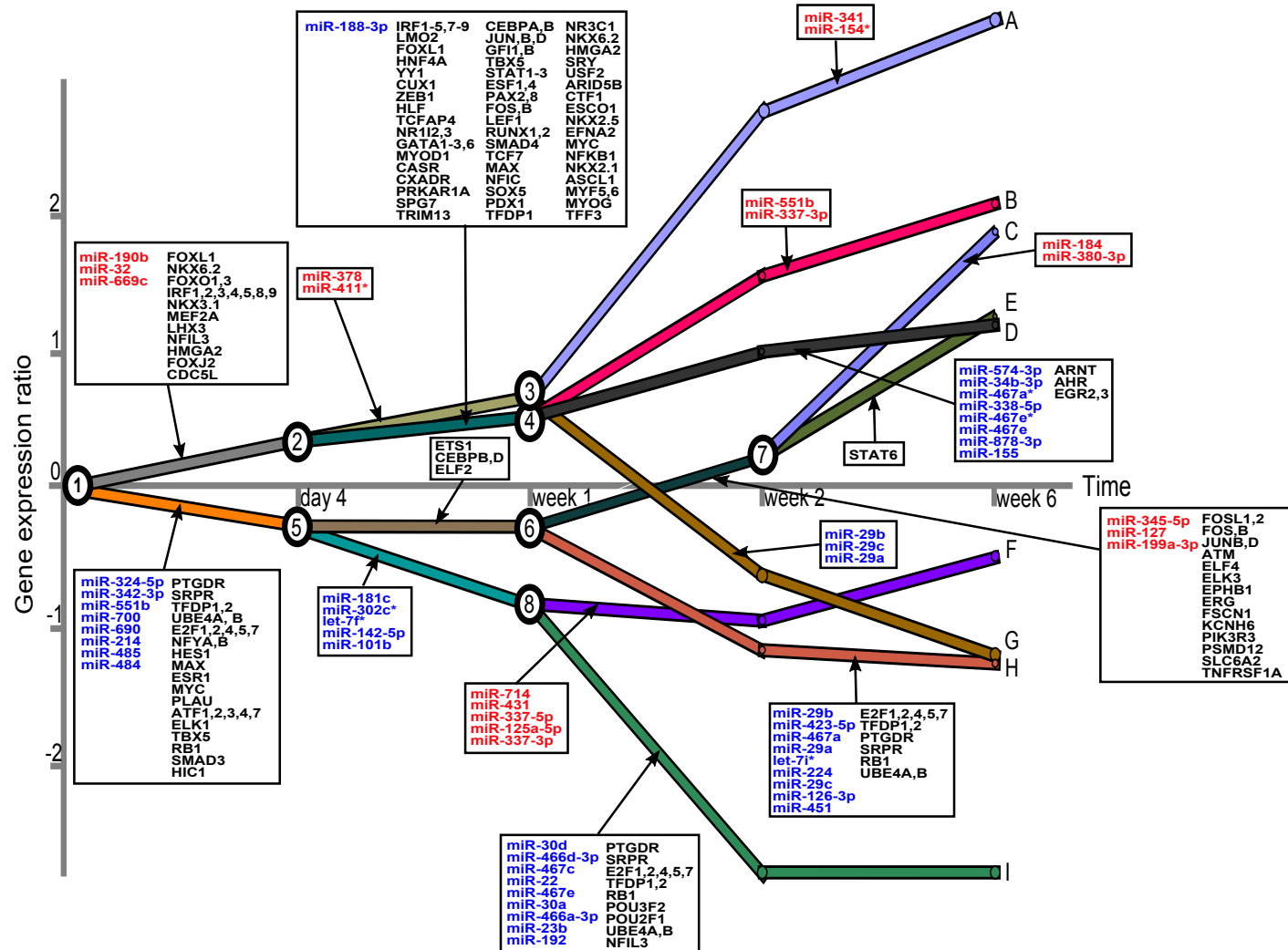
-50% death rate after 3 years



Extensive lung fibrosis
(source:wikipedia)

-pathways for lung development appear activated in reverse direction during the disease

Joint dynamic network for lung development



- 22 out of 56 miRNAs predicted by the method are differentially expressed in patient cohorts with the IPF lung disease
- different to developmental miR-30d is down regulated in IPF patients