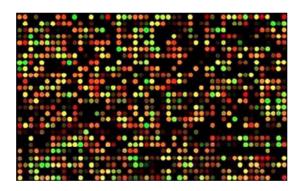
10-810: Advanced Algorithms and Models for Computational Biology

Microarrays

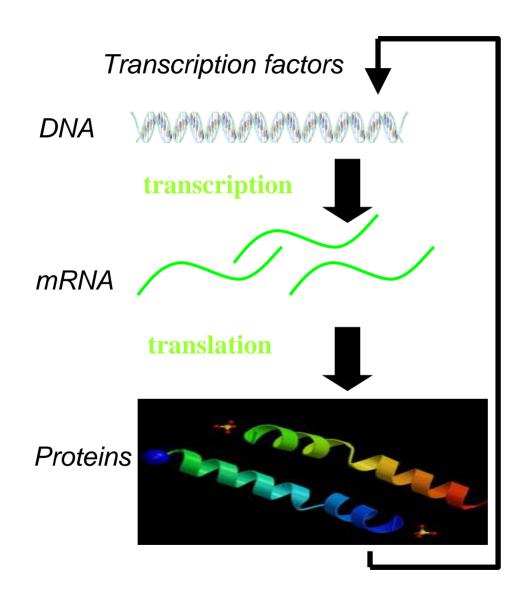


Why sequence is not enough

Identifying genes and control regions is not enough to decipher the inner workings of the cell:

- We need to determine the function of genes.
- We would like to determine which genes are activated in which cells and under which conditions.
- We would like to know the relationships between genes (protein-DNA, protein-protein interactions etc.).
- •We would like to model the various dynamic systems in the cell

Central Dogma



Genes and Gene Expression
Technology
Display of Expression Information

What is a gene?

Promoter

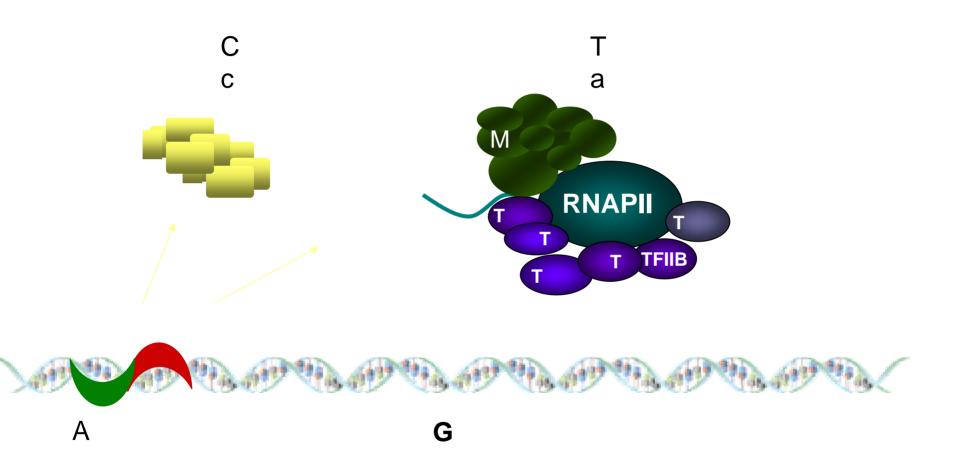
Protein coding sequence

Terminator

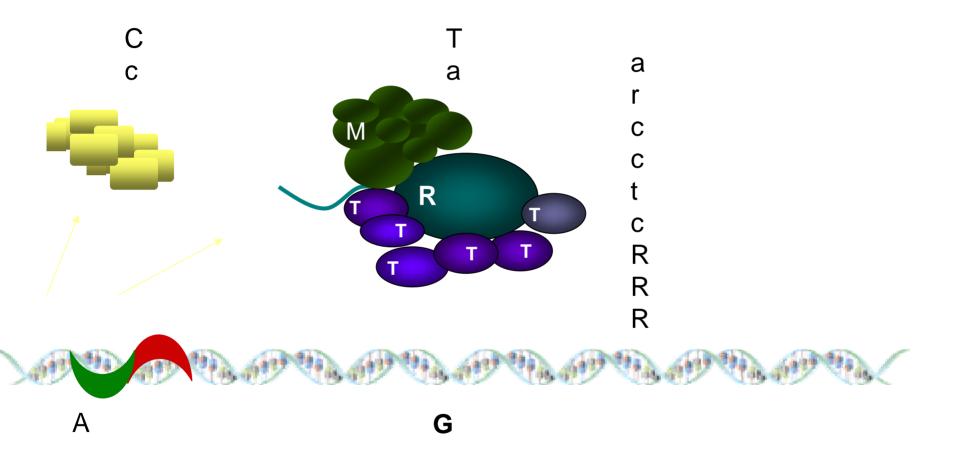


Genomic DNA

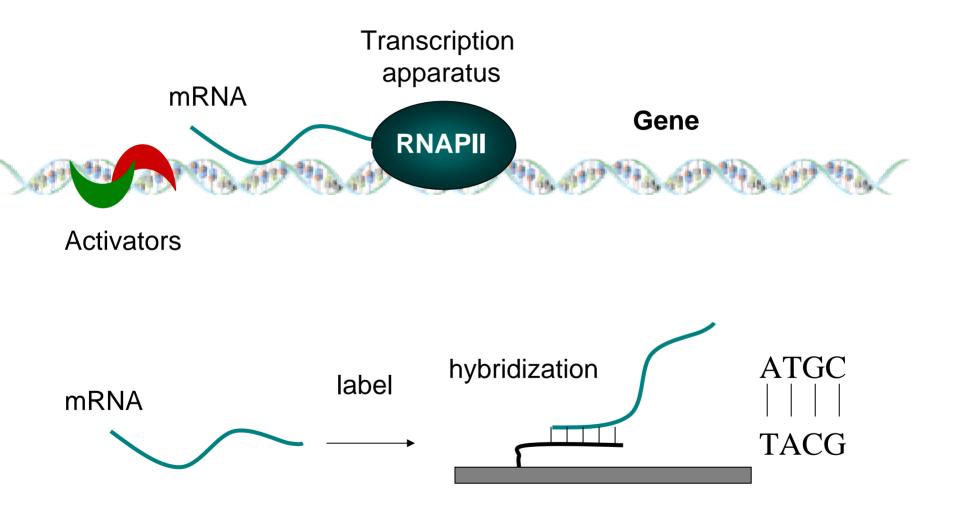
How are Genes Regulated? DNA-binding Activators Are Key To Specific Gene Expression



NA-binding activators are General Part and additional factors

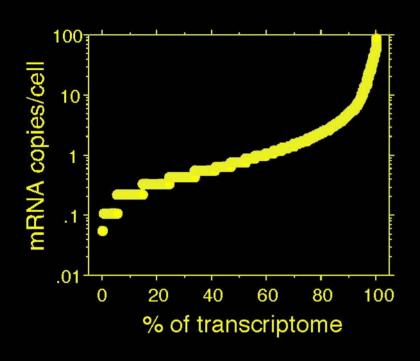


Genome-wide Gene Expression (mRNA) can be Measured with DNA Microarrays



Yeast Transcriptome (Glucose)

5460 mRNA species average level: 2.8 copies/cell median level: 0.8 copies/cell



80% of the transcriptome is expressed at 0.1 - 2 mRNA copies/cell

Some Additional Numbers

Yeast:

6200 genes

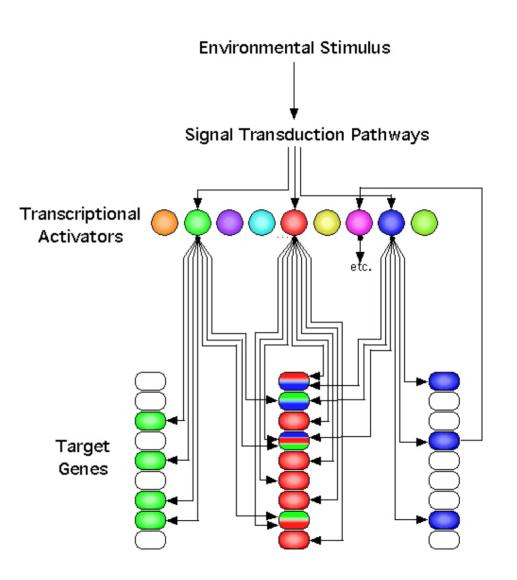
~200 transcriptional regulators

Human:

30,000 - 50,000 genes

~1700 transcriptional regulators

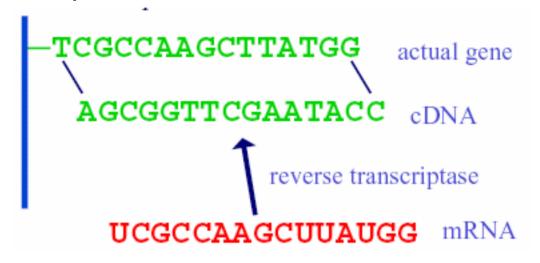
>100 cell types

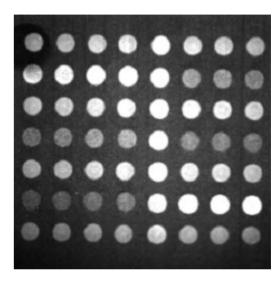


Genes and Gene Expression
Technology
Display of Expression Information

Microarray Hybridization

Watson-Crick base pairing of complementary DNA sequences.



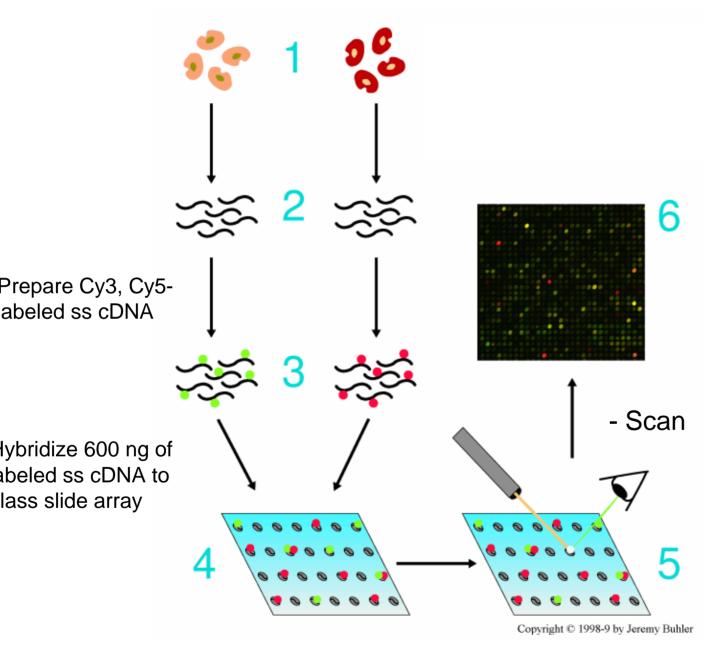


- Microarrays have thousands of spots, each representing a piece of one gene, immobilized on a glass slide.
- The intensity (or intensity ratio) of each spot indicates the amount of labeled cDNA hybridized, thus, representing the starting mRNA transcript abundance.

Two major technologies

- cDNA arrays
 - probes are placed on the slides
 - allows comparison of different cell types
- Oligonucleotide arrays
 - partial sequences are printed on the array
 - measure values in one tissue type

Trybridization and ocaliling oblivit arrays

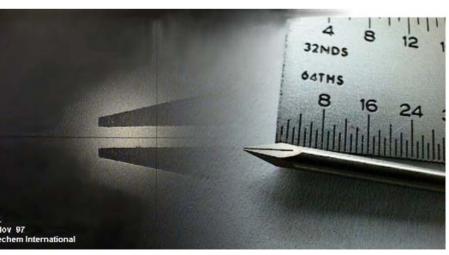


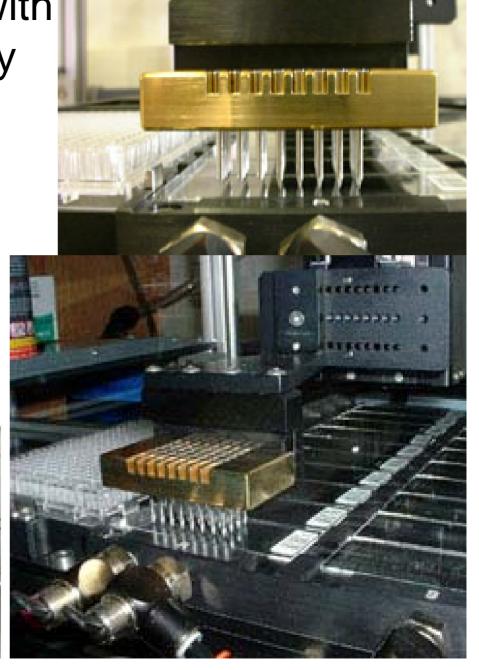
Cartesian PixSys 5500 with quill printing technology

Complete subsequences are rinted on the array 10,000 spots/slide

Spots are 100-200 µm in diameter

Hybridization volumes: 20-100ul



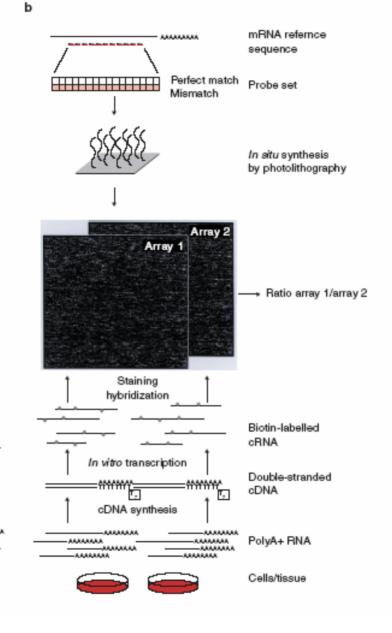


Array Scanning



Laser based - fluorescent emission

Hybridization and Scanning oligo arrays



cDNA vs. Oligo: Pros and Cons

cDNA

- Does not require sequence
- Cheap
- Direct comparisons
- Inaccurate
- Cannot measure individual samples

Oligo

- Can be designed to minimize cross hybridization
- Allows for internal control
- Both lead to better accuracy
- expensive
- limited to certain species

Errors

Microarrays introduce many errors which should be taken into account when working with measured expression values:

- Scanning errors
- Spotting errors
- Cross hybridization
- Errors related to day / reading device / experimentalist
- Background differences between slides

Error types

Microarrays introduce many types of errors which should be taken into account when working with measured expression values:

- Scanning errors additive + multiplicative
- Spotting errors multiplicative
- Cross hybridization multiplicative
- Errors related to day / reading device / experimentalist
 additive + multiplicative
- Background differences between slides additive

- Scanning errors
- Spotting errors
- Cross hybridization
- Errors related to day / reading device / experimentalist
- Background differences between slides

Analysis of image data (we assume it was performed)

- Scanning errors
- Spotting errors
- Cross hybridization
- Errors related to day / reading device / experimentalist
- Background differences between slides

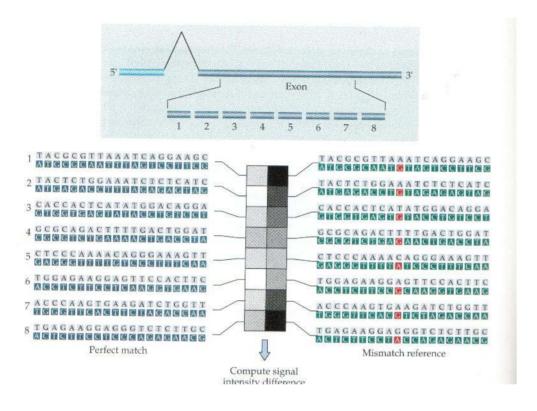
Use ratio instead of individual values:

$$Y_i = R_i / G_i$$

- Scanning errors
- Spotting errors
- Cross hybridization
- Errors related to day / reading device / experimentalist
- Background differences between slides

For Oligo arrays, use the match / mismatch spots

Match / Mismatch



- Presence and absent calls can be made using the Match / Mismatch information.
- However, it has been reported that in some cases the mismatch was higher than the match.

- Scanning errors
- Spotting errors
- Cross hybridization
- Errors related to day / reading device / experimentalist
- Background differences between slides

Normalization (next lecture)

Binding arrays

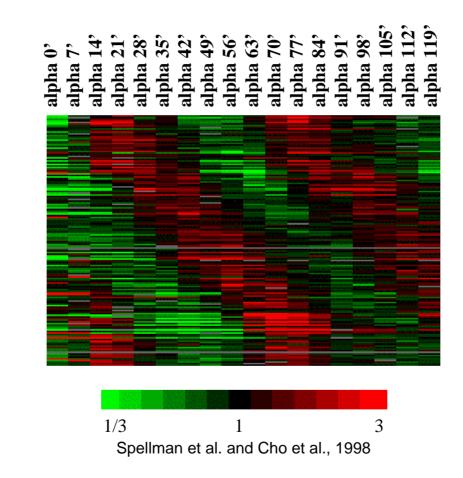
- Instead of printing the genes on the microarray, we can print the intergenic region (an area upstream of the gene).
- We tag a protein of interest (a transcription factor) and fuse all proteins to DNA.
- Next, we hybridize the extracted portions of DNA onto the array, resulting in areas that are bound by the TF being spotted on the microarray.

Genes and Gene Expression Technology

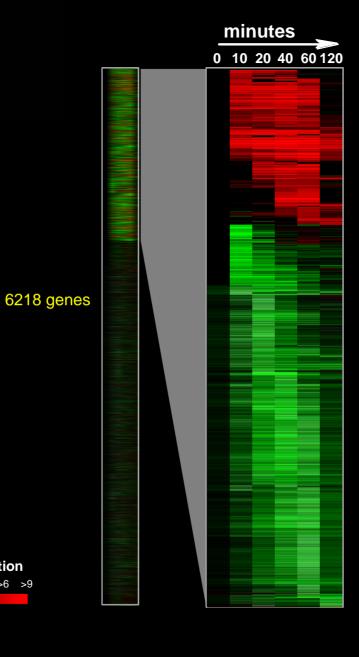
Display of Expression Information

reast cen cycle expression baseline expression program 10 20 70 80 gene 1 genes Higher Lower expression expression compared to compared to baseline baseline Spellman et al Mol. Biol. Cell 1998

Yeast Cell Cycle Gene Expression Program



800 Genes



Fold repression

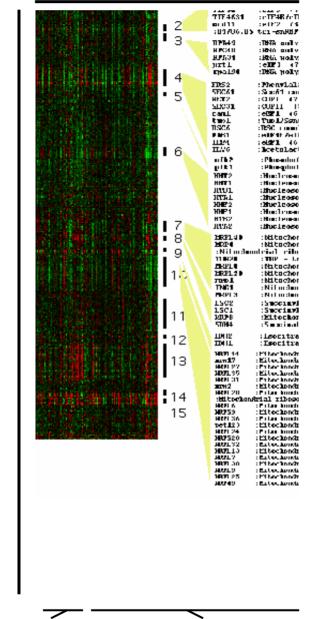
>3

>9 >6

Fold induction

>6 >9

Exercising the Genome



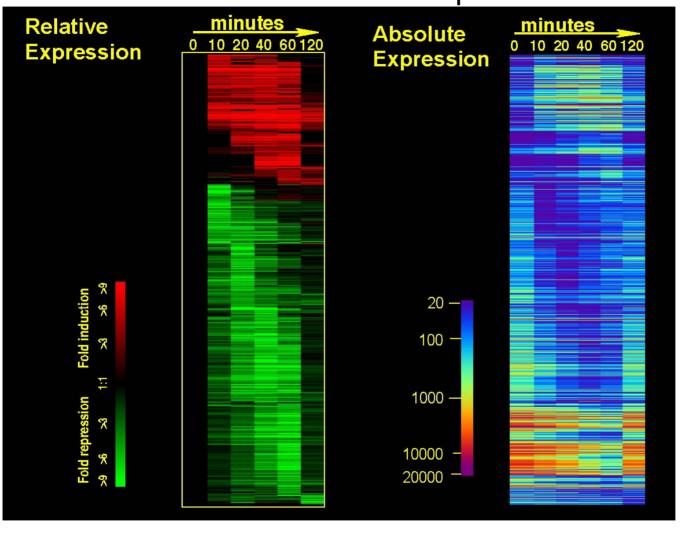
6200 Genes

Environment

Single-gene Mutations

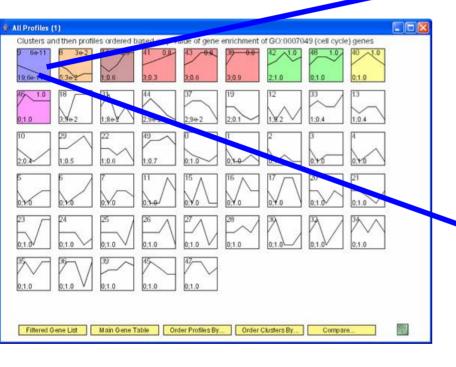
Visualization:

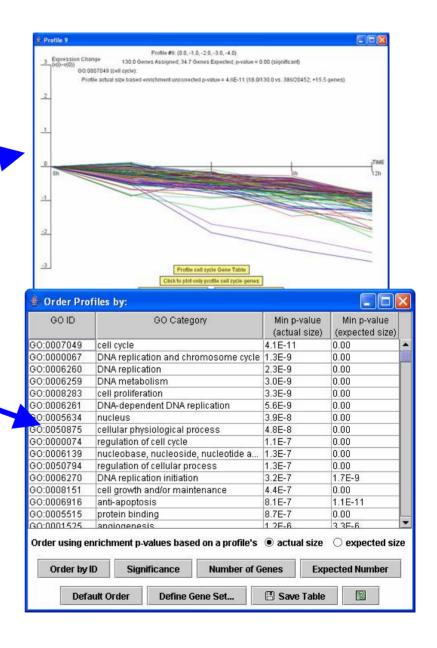
Relative vs. absolute expression

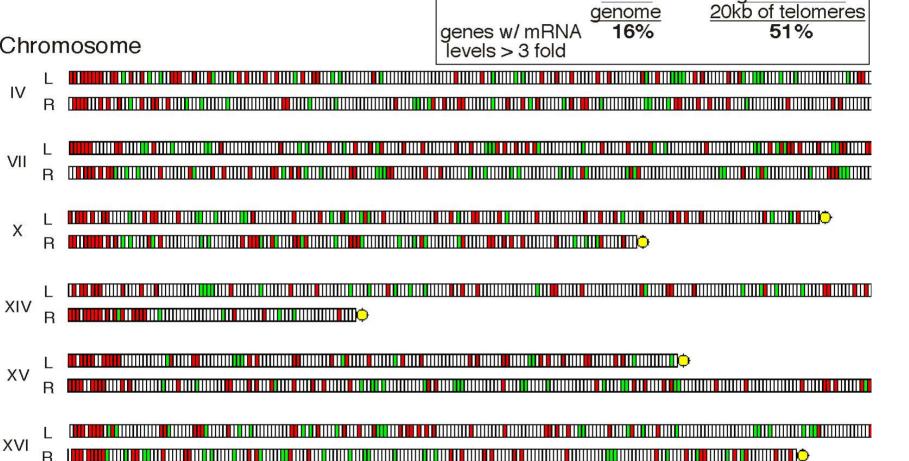


Using annotation databases

 Statistical tests to identify the overlap with various functional categories



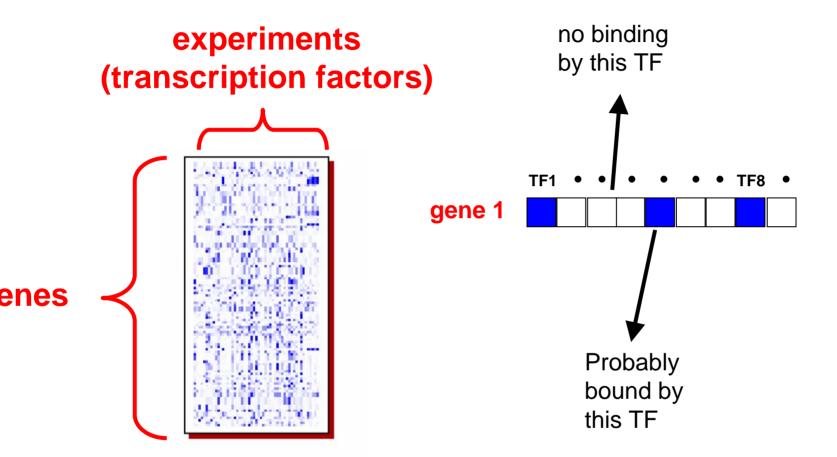




genes within

whole

Genome wide binding



What you should know

- The basic idea behind microarray profiling
- The two different microarray technologies
- Pros and cons for each
- Noise factors in microarray experiments (more next time)